Effect of Foliar Application of Yeast on Yield and Seed Quality of Some Rice Cultivars

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# ABSTRACT

Two field experiments were conducted at El-Serw Research Station, Agricultural Research Center, Egypt, during Y. YY and 2013 seasons to study the effect of 5 concentrations of dry yeast (Saccharomyces cervisia) i.e. 0, 2, 4, 6 and 8 g/L as foliar application at 25, 55 and 85 days after sowing on growth characters, yield and its components of Giza 178, Sakha 105 and Sakha 106 rice cultivars .The produced seeds obtained from the field experiment were investigated under Laboratory condition to study seed quality characters (seed, seedling vigor and its health) at Seed Technology Research Unit in Mansoura, Egypt . Results revealed that Giza 178 cv. was significantly thumping as compared with other studied cultivars in all studied characters of both growth and yield components. Maximum growth and yield components were produced from foliar spraying with 7 or ^ g/L dry yeast where show enhancement in heading date, plant height, number of tillers, and increased dray matter, number of panicles, panicle length, V... grain weight and grain yield with high significant in both seasons. The data of seed and seedling vigor test showed that Giza 1VA cv. gave the highest values of normal seedlings (GP), speed germination (SG), seedling length (SL), while Sakha ). cv. gave the highest values of dry weight followed by Sakha ). cv. as compared with other cultivars. On the other hand, foliar spraying with <sup>1</sup> or <sup>A</sup> g/L dry yeast were the best treatments of seed and seedling vigor as compared with control and other concentrations of dry yeast and also decreased abnormal seedlings and seed rot percentages with significant effect. Seed health test recorded thirteen fungus species were associated with seeds of three cultivars under study. Giza 178 cv. was the less values of total fungi, followed by Sakha V. and Sakha V. cvs. Alternaria padwickii showed maximum values presented on seeds of three cultivars followed by Bipolaris oryzae, A. longesima, Fusarium semitectum, A. tenuis, F. graminearum and Sarocladium oryzae... et. The beneficial effects of yeast lead to lower the number of seed-borne fungi with increasing concentration from 2 to ^ g/L especially pathogenic fungi which caused reduction to seed and seedling vigor of rice . It could be recommended that use of foliar spray with dry yeast (S. cervisia) at 6 - ^ g/L at 25, °° and 85 days after rice sowing gave the highest values of yield, yield components and next enhanced seed, seedling vigor and lower numbers of seed-borne fungi associated with rice seeds(seed quality).

Keywords : Rice, yeast, yield components, seed and seedling vigor, seed health

#### **INTRODUCTION**

Rice (Oryza sativa L.) is a principal cereal crop of Egypt. Foliar spraying with natural agents during different growth stages of rice plants considered an important way to enhancement vegetative growth, yield components and control of seed-borne fungi especially field fungi and subsequently production of seed related high quality. Microorganisms play an important role in affecting the quality of seed of which fungi are the largest group. These pathogens are disastrous as they reduce seed vigor and weaken the plant at its initial growth stages. The infected seeds may fail to germinate, transmit diseases from seed to seedling and from seedling to growing plants (Fakir et al., 2002). Tarp, et. al. (1987) found that Bipolaris oryza causal agent of seedling blight and brown spot disease, while Alternaria padwickii causal agent of seedling blight and stack burn disease and Fusarium moniliforme causal agent of bakane disease and foot rot. Sharma and Chahal, (1996) found negative correlation between kernel smut (Tilletia barclayana) and seed germination, seedling length and seedling dry weight, also Curvularia lunata, Drechslera tetramera and Fusarium moniliforme were predominant and caused seed rot/seedling mortality. Reves et al., (1933) reported that stunting of seedlings and a reduction in the number of tillers when smutted seeds were sown. Ram et al., (1998) stated that kernel smut of rice caused by T. barclayana was widespread and causes considerable qualitative and quantitative yield losses of rice.

Environmental conditioning and the use of chemicals to control the plant diseases are expensive remedy and may also reduce populations of beneficial microorganisms, thus the use of safe chemicals control has become more attractive (Cooke, 1993). Nowadays, a great attention has been focused on the possibility of using natural and safe agents for promoting growth of rice and for inducing its resistance against different diseases. Saccharomyces cerevisiae is considered a new promising plant growth promoting-factor for different crops. EL-Sirafy, et al., (2011) found that inoculation with biofertilizer mixtures along with foliar spray with yeast or yeast + N- solution lead to the highest grain yields, straw yields, weight per plant, dry matter plant and could be used along with application of 50 to 75 % of the recommended rate of N, thus saving 25 - 50 % of N requirement of rice. Mady, (2009) found that foliar application with yeast extract increased many growth aspects as number of leaves per plant, dry weights of both stems and leaves and total leaf area, also increased photosynthetic pigments, total sugars, free amino acids and crude protein content in leaves of faba bean. Yeast treatments were suggested to play a beneficial role in cell division and cell enlargement (Natio, et al. (1981). Yeast as a natural stimulator is also characterized by richness in protein, carbohydrates, nucleic acid, lipids and different minerals and Li in addition to thiamin, riboflavin, pyridoxine, hormones and other growth regulating substances, biotin, B12 and folic acid (Nagodawithana, 1991). Shalaby and El-Nady (2008) found that seed soaking, foliar spraying or soil



inoculation with S. cerevisiae increased germination rate, survival of sugar beet plants and reduction of pre-, post emergence damping off and inhibited Fusarium oxysporum liner growth in Lab. studies. El-Emery (2004) reported that germination rate of different seeds of barley, maize, pea and been was great enhanced by using different yeast dilutions and growth of plumules, root lets and cotyledon enlargement was also stimulated. Elwakil, et al., (2009) showed that the yeast was effective in reducing the liner growth of Cephalosporium sp., F. oxysporum and F. solani, also they found that pre- and post- emergence damping off was reduced significantly when seeds of faba bean were coated with a water suspension of the yeast  $(10^9 \text{ cfu})$  $mL^{-1}$ ). Mohamed, et al., (2015) showed that foliar application with yeast extract significantly increased all investigated morphological characters of vegetative growth and yield, also induced favorable enhancements in most of included tissues of the main stem and leaves especially in conducting tissues (phloem and xylem) of Basil plant (Ocimum basilicum L.). Abdel-Hakim et al., (2012) revealed that foliar application with different concentrations of yeast enhanced growth, yield and yield components along with the chemical composition of fresh and/or dry plants and green pods of snap bean (Phaseolus vulgaris L.). Said, et al., (2012) reported that foliar application with yeast increased significantly the vegetative growth, yield and its quality of cucumber. Amer, (2004) indicated that the yeast is a natural source of cytokinins and has stimulatory effects on bean plants. Yeast extract was suggested to participate in a beneficial role during vegetative and reproductive growth through improving flower formation and their set in some plants due to its high auxin and cytokinins content and enhancement carbohydrates accumulation (Barnett, et al., 1990). In this connection, yeasts have been reported to rich source of phytohormones (cytokinins), vitamins, enzymes, amino acids and minerals (Fathy and Farid, 1996; Khedr and Farid, 2000 and Mahmoud, 2001). Darweesh, et al., (2003) reported that yeast contains tryptophane which is the precursor of indol acetic acid which promotes plant growth. The present study amid to evaluate foliar application with yeast (Saccharomyces cerevisiae) and its effect on characteristics of growth yield, its components, seed and seedling vigor and its health of some rice cultivars.

#### **MATERIAILS AND METHODS**

This investigation was carried out in El-Serw Agricultural Research Station Farm, ARC during two successive summer seasons of  $(\cdot)^{\uparrow}$  and 2013 to study the effects of foliar application of dry yeast (*Saccharomyces cerevisiae*) was identified in the Microbiology Department, Facility of Agriculture, Mansura University on growth characters , yield and its components of three rice cultivars known as; Giza 178, Sakha  $(\cdot)^{\circ}$  and Sakha  $(\cdot)^{\neg}$  was used .Yeast was applied as foliar application with five concentrations *i.e* 0,2,4,6 and 8 g/L.

The experiment were arranged in strip plot design with four replicates. The vertical plots were denoted for rice cultivars, while the horizontal plots were arranged for yeast treatments. The area of each experimental plot  $(2\times5 \text{ m}^2)$ . Rice grain of the three cultivars at rate of 60 kg/ fed. were soaked in fresh water for 24 h and incubated for 48 h for pre-germination, the seeds were sown directly (Broadcast) at  $25^{\text{th}}$  May in the first and second seasons. The foliar spraying treatments of yeast were prepared by dissolved active dry yeast in water, followed by adding sugar at ratio 1 : 1 and kept for two days for activation and reproduction of yeast before application on the plants. The control plants were sprayed with tap water. The foliar spraying was repeated three times *i.e.* 25, 55 and 85 days after sowing for each concentration. The chemical fertilizers and weed herbicide were added as recommended doses . **Studied characters:-**

Heading date was recorded at following time for each cultivar, as the number of days from the date of up to 50% heading. At harvest, plant height was recorded for every plot, number of tiller  $/m^2$  was counted, number of panicles/ $m^2$ , dried and weighted as dry matter production g/ $m^2$ . Ten random panicles were collected from each plot for estimating panicle length, number of filled grains /panicle and weight, number of unfilled grains/panicle and 1000 grain weight. Ten guarded square matters were harvested for grain, straw yield estimation, grain yield was adjusted to 14% moisture content.

# Quality of seed production test

#### Seed vigor test :-

Rice seed samples of field experiment for all treatments of the two seasons were collected and transferred to investigate in Lab. of Seed Technology Research Unit, Mansoura, Egypt. Seeds were sown on Petri-dishes (12 cm) contained three layers of moistened blotters, eight replicates of 25 seeds/Petri (200 seeds) from each treatment and incubated in the growth chamber (Seedburo Equipment company, USA) for 14 days night lengths 15/9 h at 25 °C  $\pm$  2 and evaluated the following characters :-

 Mean germination percentage (MGP %) (normal seedlings) it was calculated by counting only normal seedlings (ISTA, rules 1999).

$$MGP = \frac{n1 - n2}{n1} \times 100$$

The mean germination percentage (MGP) . , n1 is total number of treated seed plated, n2 the number of abnormal seedlings plus seed rot.

2 -Speed germination index (SGI): It was calculated as described in the (AOSA, 1983) by the following formula :

- 3- At the final count, five normal seedlings from each replicate were randomly taken to measure seedling characters, seedling length (cm), dry weight (g) of seedlings (ISTA rules ,1999).
- 4- Seedling vigor index (SVI) = MGB × seedling dry weight Seed health testing by blotter method :-

Two hundred rice seeds of each cv. and treatment were planted on three layers of filter paper will soaked in distilled sterilized water in eight replicates (25 seeds / Petri dish 9 cm). Another two hundred un treated seeds as a control were incubated at 25  $^{\circ}C \pm 2$  for seven days under alternating cycles of 12 hours near ultraviolet (NUV) light and darkens. Percentage of fungi under investigation were calculated by stereo-binocular microscope (Wild Heerbrugg 6.3 - 32x), whenever necessary the compound microscope (Jenaval 3.2 -100x) was used for details and confirmation. Seed-borne fungi were identified morphologically on the basis of colony color, shape and size, mycelial growth, spore shape and size as described in the technical bulletin on seed-borne diseases and seed health testing (Chidambaram, 1973; Ellis, 1980; Booth, 1985; Burgess et al. 1988; Agarwal et al., 1989) and recorded to the following formula:-

Fungal % = 
$$\frac{n1}{n2} \times 100$$

 $n1\ the\ seeds$  with fungal growth . ,  $n2\ the\ number\ of\ treated\ seeds.$ 

Seed health testing for kernel smut (Tilletia baraclayana):-

Soaking 2000 seeds from each treatment and cultivar in two replicates of 1000 each in 0.2 % solution of sodium hydroxide (NaOH) for 24 h at 18-25 °C. Decant the solution ; spread the swollen seeds on a filter paper. Examine the seeds visually. Infected seeds will show shiny, jet-black discoloration while seeds with only brown or dull black discoloration are not infected. Bunt spores will be released from the infected, shiny,

jet-black seeds, while no spores will come out form healthy seeds or those with dull black discoloration. The number of infected seeds was recorded and percentage of infection was calculated (Agarwal & Srivastava, 1981, 1985).

### Statistical analysis :-

Collected data for each seasons were statistically analyzed by the technique analyses of variance for strip plot design and the least significant differences ( L S D ) was used to test the differences among treatments (Gomez and Gomez ,1984). Bartlett test was done to the homogeneity of error variances. The test was significant for all field traits, while was in significant for all Lab. traits, thus the data of both seasons were combined .

# **RESULTS AND DISCUSSION**

Results of growth characteristics, yield and its components of the three rice cultivars as affected by application of different concentrations of yeast are presented in Tables 1 and 2 Rice cultivars were significantly varied among them in the two seasons regarding with all characters under study. Giza 178 cv. produced the highest values of number of tillers (m<sup>2</sup>), dry matter (g/m<sup>2</sup>), number of panicles (m<sup>2</sup>), panicle length (cm), number of filled grains/panicle, panicle weight (g), grain yield (t/ha) and straw yield (t/ha), followed by Sakha 106 cv., while Sakha105 gave the highest values of 1000 grain weight (g) in both seasons compared with other cultivars .

Charactera	Heading date (day)		Plant h	eight (cm)	Number of t	illers (m <sup>2</sup> )	Dry mat	ter (g/m <sup>2</sup> )	N. of par	nicles (m <sup>2</sup> )	Panicle le	ength (cm)
Characters	2.12	7.14	2.12	1.14	2 . 1 2	1.17	1.11	1.17	7.17	2.12	2.12	7 . 1 7
					Cu	ltivars:						
Giza 178	90	٩٨	٩٤٧	٨٥٩	201	3777	٥٩٧	٤٨٧	٤١٧	371	Y.77	۲۰,٤
Sakha 105	$\Lambda\Lambda$	٨٩	97.7	Λź Λ	٤١.	317	072	۳٩.	372	292	19.7	14.5
Sakha 106	٩٢	٩ ٤	90.7	AV.V	510	371	०२१	٤ • ٢	340	٣. ٤	14.5	17.0
F test	*	*	*	*	*	*	*	*	*	*	*	*
L S D 5%	١.•	۲. •	١.٤	١,٦	37	۲۷	17	٤٣	۲۸	۲.	• . ٤	1.1
					Yeast 7	Freatments	:					
Control	۹١	٩٣	95.7	Λź Λ	341	310	٤٩٧	٤٠٩	30 M	۲۹۸	19.0	14.7
۲g/L	۹١	٩٤	95.1	10.1	٤٠٩	372	٥٣٣	519	371	317	19.1	14.4
٤g/L	۹١	٩ ٤	95.5	Λ٦.٢	٤ ٢ ٤	322	٥٧.	٤٢ ٤	377	370	19.1	14.
٦g/L	۹١	٩٤	95.7	Λ٦ ۸	えそり	307	٦.0	٤٣٨	392	222	۲۰.۰	14.
^g/L	٩٢	٩ ٤	90.0	٨٨. •	٤٦٣	301	777	220	580	377	۲. ٤	11.0
F test	*	*	*	*	*	*	*	*	*	*	*	*
L S D 5%	١	١	٥.١	1.4	۳١	1 Y	۲٤	30	27	١٨	•.0	•.٧
Interaction	NS	NS	NS	NS	*	*	NS	NS	*	*	NS	NS

 Table 1: Effect of application of different yeast concentrations on heading date, plant height, number of tillers, dry matter, number of panicles and panicle length of some rice cultivars in the two seasons

Regarding to the effect of yeast concentrations on the studied characteristics, data in Tables 1 and 2 indicated that foliar application of *S. cervisia* at 8g/L gave maximum values of number of panicles ( $m^2$ ), 1000 grain weight (g) and grain yield (t/ha) as (435), (26.3) and (6.0), respectively in the first season, followed by 6 g/L then 4 g/L compared to other concentrations and control. Increased foliar application of yeast concentrations from 2 to 8 g/L gave enhancement of growth characters and yield components with different significant for all characteristics under study in both seasons. Similar results were reported by Natio, *et. al.*, (1981) and Mady, (2009). El-Ghamriny, *et. al.* (1999) found that dry bread yeast (*S. cerevisiae*) is a kind of the used biofertilizers in soil fertilization or in foliar application on the shoots of vegetable crops. This is may be due to it's content of many nutrient elements and being productive of semi growth regulator compounds like auxins, gibberellins and cytokinins, the positive effects caused by the addition of yeast suspension in improving shoot characteristics might be due to direct or indirect effect of the yeast throughout its ability in changing the environment of roots or because the development of the yeast after analysis into wide groups of amino acids and vitamins (Glick, 1995).

	N. of	filled	Panicle	weight	N. of u	infilled	1000-gra	in weight	Grai	Grain yield S		Straw yield	
Characters:	grains/P		(	(g)	. ;	g/p	(	g)	(t/	ha)	(t/	ha)	
	4.14	2.12	2.12	7.14	1.11	7.17	1.11	7.18	1012	1.14	1.11	1.17	
					Cultiv	vars:							
Giza 178	157	1.1	۳.•	۲. •	٨	١٣	۲١,٦	۲۰.۱	٦.٨	0.1	٨.٨	٥٩	
Sakha 105	77	۲۷	۲.٤	١.٩	10	10	۲۸.0	۲٦.٤	٤٧	٣.٢	٧.٤	۰. •	
Sakha 106	۱ • ٤	~~	۲.۲	1.7	۱.	۱۳	۲۷.٤	25.1	٥٣	٣.٦	٧.٩	٥٤	
F test	*	*	*	*	*	*	*	*	*	*	*	*	
L S D 5%	٤	٩	• . ٢	• ٢	٩	١	•.٧	1.1	•.0	• . ٢	٠٤	٠٦	
					Yeast Tre	atments:							
Control	1.7	~~	۲٫۳	1.4	١٢	10	10.1	۲۳.۰	0.1	٣.٦	٧.0	0.1	
۲g/L	117	٨١	٢٥	1.1	١٢	10	10.1	17.1	ه ه	۳.۸	٧.٩	0,1	
٤g/L	۱۱۳	۸۳	۲.0	1.9	١٢	١٤	10.V	۲۳.۹	ه ه	٤.•	٨.•	0 2	
٦g/L	115	۸۳	۲٦	۲. •	۱.	١٢	۲٦.٢	۲۳.۷	0.9	٤.•	٨٣	0.9	
^g/L	115	٩٢	۲.٧	۲. •	۱.	١٣	۲٦٫٣	۲٤. •	٦.•	٤٣	٨٤	٦.٠	
Ftest	*	*	*	*	*	*	*	*	*	*	*	*	
L S D 5%	٥	٦	• ٢	• ٢	١	١	٠٦	• ^	• .٣	• . ٢	٠٦	٠٦	
Interaction	NS	NS	NS	NS	NS	NS	*	*	NS	NS	NS	NS	

Table 2: Effect of application of different yeast concentrations on n. of filled grains/p., n. of u	nfilled g/p.,	panicle
weight, 1000-grain weight, grain vield and straw vield of some rice cultivars in the	two seasons	

The results in Table 3 indicated that interaction between cultivars and yeast concentrations had significant effect on number of tillers (m<sup>2</sup>), number of panicles (m<sup>2</sup>), 1000-grain weight (g) and grain yield (t/ha) of three cultivars under study, the treatment with foliar application 8 gm/L was the best of concentrations, which gave the highest values followed by 6 gm/L with all characters under study in both seasons. These results are in agreement with those obtained by (Barnett, et. al., 1990; Amer, 2004; El-Sirafy, et. al., 2011; Said, et. al., 2012 and Mohamed, et. al., 2015). The major effect of foliar spray of yeast might be due to that yeast induces nutrient minerals absorption through general improvement due to the ability of yeast to increase the production of stimulants for plant growth, especially gibberellins, auxins and cytokinins which act to improve the plant cell division and its growth (El-Bassiony, et.

al.,2014). Also, Ghoname, et. al. (2010) found that promoted plant vegetative growth i.e. plant height, number of leaves and branches, fresh and dry weight, compared to control by used yeast (1, 2 and 3 g/L) as sprayed on sweet pepper. Rania, et. al., (2011) found that foliar application with relatively used concentration 50 and 100 ml/L induced significant promotive effects on all investigated morphological plant characters (plant height, number of branches/plant, number of leaves/plant, total leaf area/plant and shoot dry weight/plant) and yield components of kidney bean. Abdel-Hakim, et. al., (2012) showed that spraying the snap bean plants with yeast in different concentrations enhanced plant growth and most of the studied characteristics and reported that yeasts is a rich source of phytohormones (especially cytokinins) vitamins, enzymes, amino acids and minerals .

 Table 3: Means of number of tillers, number of panicles, 1000-grain weight and grain yield as affected by the interaction between cultivars and yeast concentrations during two seasons .

Caltingang	Yeast	Number of	tillers (m <sup>2</sup> )	Number of pa	anicles (m <sup>2</sup> )	1000-grain	weight (g)	Grain y	ield (t/ha)
Cultivars	Treatments	2.12	۲۰۱۳	۲۰۱۲ -	۲۰۱۳	۲۰۱۲	۲۰۱۳	۲۰۱۲ آ	۲.۱۳
	Control	٤٠٦	301	۳۸۸	٣٤.	۲۰٫٦	19.7	٦٣	٤٧
	2 g/L	٤٢٨	۳۷.	397	37V	Y1.Y	19.2	٦.٨	٤٩
Giza 178	4 g/L	207	۳۷۹	٤١٨	317	111	۲۰.٩	٦.٨	۰. ۰
	6 g/L	٤٨٤	٤•١	209	377	177	۲۰٫۱	٦٩	0.2
	8 g/L	572	٤.٢	397	۳۹.	177	۲۰.٤	٧.١	0.2
	Control	٣٤.	292	222	200	7 Y A	10.7	٤.•	۲۸
	2 g/L	٤•٤	T1V	٣٣٤	787	Y A. Y	10.V	٤٢	۳.۱
Sakha 105	4 g/L	٤١٤	T1V	r7r	262	۲٨ ٤	۲٦.٤	٤٠٤	۳.۱
	6 g/L	٤٣٠	379	۳۸۸	3.5	۲٨٦	٥.٢٢	٥.٣	۳.0
	8 g/L	587	۳۳.	٤.0	۳.0	۲٨.٨	11.11	0.0	۳.٧
	Control	٣٩٦	۲٩.	305	219	۲٦٫٨	17 V	۰. •	٣٣
	2 g/L	٣٩٦	315	5 V Y	۳	۲٦ <sub>.</sub> ٩	15.1	0.7	٣.٤
Sakha 106	4 g/L	٤.0	۳۳.	344	511	۲٧.٦	٢٤.٣	0.7	٣.٧
	6 g/L	٤١٠	377V	۳۸.	311	7 Y A	٢٤.٣	०,٦	٤.•
	8 g/L	٤٦٨	٣٤.	٤٤.	۳۲.	۲٧.٩	15.1	०,٦	٤.•
L S D 5%	C	0 5	29	٤٤	۳۱	1.1	1.7	•.0	• . ٣

### Quality of seed production from field experiment :-Seed and seedling vigor :-

Table 4 shows the effects of cultivars and yeast concentrations on seed and seedlings vigor, Giza 178 cv. recorded the highest percentage of normal seedlings (germination percentage) (85%), speed germination (SG) (27.8), seedling length (11) cm and less value of seed rot (6%), dry weight (0.18) g, followed by Sakha

105 and 106 cvs with high significant differences for the studied characters between cultivars. The effect of foliar spray of yeast was statistically significant for normal and abnormal seedlings, seed rot, speed germination, seedling length and dry weight as seed and seedling vigor, sprayed at 8 g/L gave the highest values of normal seedlings (91%), lower abnormal seedlings (6%), seed rot (3%), (27.7) speed germination, (0.23) g

dry weight while 6 g/L gave (11) cm seedling length as compared with control and other concentrations. Perceiver from the treatments that increased foliar spraying of concentrations with yeast lead to improved of vigor characters of seed and seedling of rice, also the interaction of cultivars and treatments was found statistically significant in case of normal, abnormal seedlings and dry weight. This was in agreement with the data obtained by Muller and Leopold, (1966) who demonstrated that enhancing effect of yeast application might be due to yeast cytokinins enhancing the accumulation of soluble metabolites. Also, the enhancing effect of yeast on germination rate and on the vegetative growth parameters was strongly supported by Entian and Fröhlich (1984), they stated that increased enzyme activity regulating catabolic productions in eukaryotic cells treated with *S* . *cerevisiae*. El-Emery (2004) who reported that germination rate of different seeds of barley, maize, pea and bean was greatly enhanced by using different yeast dilutions and growth of plumules, rootlets and cotyledon enlargement was also stimulated. Aml and Abd El-Hai (2011) showed that peanut seed treatments with *S* . *cerevisiae* improved germination criteria, seed quality, morphological and physiological characters of seedlings .

Table 4: Effect of application of different yeast concentrations on seed and seedlings vigor of some rice cultivars .

Characters	+ Normai seedings	++ Abnormal seedings	+++ Seed rot	Speed germination	Seeding length	Dry weight
		Cul	tivars (A)			
Giza 178	٨٥	٩	٦	Y. YY	11.0	• 14
Sakha 105	٨٣	٩	Α	۲٦,٨	۸.۳	. 10
Sakha 106	٨٣	v	۱.	4.77	٩.٢	• . ٢ •
F test	*	*	*	NS	*	*
L S D 5%	١	Ŋ	۲	-	۱.۰	• • • •
		Yeast T	reatments (B)			
Control	٧٥	1 £	11	۲۳.۰	۲.٦	• 14
2 g/L	7	٩	٩	۲٥	٨٩	0.20
4 g/L	٨٣	Y	۱.	Y 0.Y	٩.٤	• 11
6 g/L	٨٦	Y	v	Y.Y7	11.	• 77
8 g/L	91	٦	٣	Y.Y7	۱۰.٦	• . ٢٣
F test	*	*	*	NS	*	*
L S D 5%	۲	٣	٣	-	1.0	• • • •
Interaction A×B	٤	٤	NS	NS	NS	• • • ٢
			1 111 5			

The results gave evidence of infection by seedborne fungi Table 5 thirteen fungal species among the tested three rice cultivars; Alternaria padwickii was the highest percentage on seeds of Sakha 105 cv. with (20.5%) followed by Giza 178 cv. (12%) thin Sakha 106 cv. (11%) with high significant differences. Bipolaris oryza recorded on S. 105 cv. seeds the highest value (19.7%) followed by Sakha 106 cv. (9%) and Giza 178 cv. (8.8%) with high significant. Fusarium graminearum was (4.5%) for each Sakha 105 cv. and Giza 178 cv. seeds. F. moniliforme was (2.1%) for Sakha105 cv. followed by Sakha 106 (1.3%) and (0.8%) with Giza 178 cv. of seeds. Sarocladium orvzae recorded (3.7%) on Sakha 106 cv., (2.1%) on Giza 178 cv. and (1.6%) on Sakha 105cv. seeds. Tilletia barclayana was for Sakha 105 cv. (1.1%), Giza 178 cv. (0.7%) and (0.3%) on Sakha 106 cv. seeds with high significant . Sakha 105 cv. recorded the highest total fungi on seeds was (77.1) followed by Sakha 106cv. (49), while Giza 178 cv. was the lowest (45.4) as compared of cultivars.

Effect of yeast concentrations on frequency of seed borne- fungi, the results showed that *Alternaria longesima* decreased from (15%) as control to (0.4%) with 8 g/L as concentration, *A. padwickii* recorded the highest percentage of infected seeds (26.2%) as control and decreased to (4.9%) with 8 g/L yeast, *B. oryza* was (20.4%) in control and reduced to (4%) with 8 g/L dry yeast.

*Fusarium semitectum* was decreased from (11%) to (0.4%), *F. graminearum* was from (7.1%) to (%·.<sup>2</sup>) and *F. moniliforme* was from (3.1%) to (0.4%), *Sarocladium* 

oryzae from (5.3%) to (0.0%), *Curvularia oryzae* from (3.6%) to (0.0%), *Tillieta barclayana* from (0.7%) to (0.2%).Total fungi decreased with increasing concentrations of yeast from (113.9) as control to (82.3) with 2 g/L, to (51) with 4 g/L, to (24.9) with  $^{\intercal}$  g/L, to (11.6) with 8 g/L as foliar spray. The observed increasing of yeast concentrations (2, 4,  $^{\intercal}$  and 8 g/L) leads to increasing reduction of the disease incidence with high significant of all fungi except *Cladosporium* sp. and *F. moniliforme*.

Data Table 6 show that A. padwickii recorded the highest value on Sakha 105, Giza 178 and Sakha 106 cvs by (36), (21.3) and (21.3%) respectively, followed by B. oryza with (29.3), (17.3) and (16%) on Sakha 105. Sakha 106 and Giza 178 cvs seeds respectively, A. longesima recorded on seeds Sakha 105 cv.(22.6%), Sakha 106 cv. (13.3%) and Giza 178 cv. (9.3%), F. semitectum was (13.3%) on Sakha 105 cv., (10.6%) on Giza 178cv. and (9.3%) on Sakha 106 cv. seeds, F. graminearum recorded (8%) on each Giza 178 and Sakha 106 cvs and (5.3%) on Sakha 105 cv. seeds. Fusarium moniliforme, S. oryzae and T. barclayana recorded on Giza 178 cv. (1.3), (4) and (1%), respectively for the control. On the other hand, the data showed that seed treatment with different from yeast concentrations lowered in general number of seed- borne fungi with increasing the concentrations with all cultivars, where A. padwickii, B. oryzae, F. graminearum and T. barclayana were decreased to (5.3), (2.7), (0.0) and (0.2 %) on seeds of Giza 178cv., (5.3), (8.0), (1.3) and (0.3%) for Sakha 105 cv. and (4.0), (1.3), (0.0) and (0.1%) for

Sakha 106 cv. seeds respectively with treatment 8 g/L. *A. padwickii* was the most spread on seeds of three rice cvs under study which recorded (221.2) as total fungi followed by *B. oryzae* (187.9), *A. longesima* (112.5), *F. semitectum* (71.6), *A. tenuis*, (65.9), *F. graminearum* (65.1), *S. oryza* 

(37.1) and *Curvularia oryzae* were (26.5), *Cladosporium* sp. (24), *F. moniliforme* (21.1), *Nigrospora oryzae* (14.6), *Stemphylium botryosum* (10.6) and *T. barclayana* (7.5) respectively.

 Table 5: Effect of application of different yeast concentrations on frequency of seed- borne fungi of rice by

 Blotter method .

· <b>-</b> .	ра	skii	5		ium.	ria	unu	əm.	un	ora	0	ium m	ı ma	=
Fung	A. longesi	A .padwic	A . tenui	B. oryza	Cladospor sp.	Curvula oryza	F. graminea	F. monilifor	F. semitect	Nigrospo oryzao	S. oryza	Stemphyl botryosi	Tilleti barclaya	Total Fung
						Cult	ivars:							
Giza 178	۳.0	17.0	٥٩	٨.٨	۲.۱	• .0	٤٥	• ^	۳.٧	• ^	۲.١	•.•	•. ٧	20.2
Sakha 105	17.0	۰.٥	٤.٠	19.7	۳.۱	۲.۷	٤٥	۲.١	0.1	•.٧	٦١	1.1	١٫١	٧٧.١
Sakha ۱۰٦	۰. •	11.0	٣.٥	٩.٠	1.7	۲.١	٤.٠	1.1	٤٨	1.9	٣.٧	1.1	• ٣	٤٩.٠
F test	*	*	NS	*	NS	NS	NS	NS	NS	*	*	NS	*	-
L S D 5%	۲.٤	۳. •	-	٣.٨	-	-	-	-	-	۲.۱	١.٧	-	• ٢	-
						Yeast Tr	eatments	:						
Control	10.	۲٦ ۲	17.0	۲۰,٤	۳.١	٣٦	٧.١	۳.١	11.0	٣.٦	0.7	۲٦	•. ٧	115.7
۲g/L	1.1	19.0	٤٠٤	11.	2.7	۳.۱	٦٧	۲ ۲	٥٣	1.7	٤٤	• . ٤	۰٫٦	11.71
٤g/L	٤٤	١٤.٠	٣٦	11.0	1.4	1.1	0.V	• ^	٤٤	•.•	۲٦	• . ٤	۰,٦	01.0
٦g/L	٤٠	٨.٨	1.1	٤٨	• . ٤	• 1	1.4	• . ٤	۲.٧	•.•	•.•	•.•	• . ٣	٢٤.٩
Åg/L	• . ٤	٤٩	٩	٤٠	• •	• •	• . ٤	• . ٤	• . ٤	•.•	• •	• .•	۰.۲	11.7
Ftest	*	*	*	*	NS	*	*	NS	*	*	*	*	*	-
L S D 5%	۳.۱	٣٩	٣٢	٣٣	-	۲_٤	۳ <sub>.</sub> .	-	٣٩	١,٦	۲.۲	٩.٥	۰.۲	-

 Table 6: Interaction between cultivars, concentrations of yeast and the effect on percentages of seed-borne fungi associated of some rice cultivars.

		Fungi												
Cultivars	Treatments	A. longesima	A Padwickii	A . tenuis	B. oryza	Cladosporium sp.	Curvularia oryza	F. graminearum	F. moniliforme	F. semitectum	Nigrospora oryzae	S. oryzae	S. botryosum	Tilletia barclayana
	control	٩٣	۳.۲۲	17.0	١٦.٠	٥٣	۱.۳	٨.٠	١,٣	۲۰٫۲	۲.۷	٤.٠	•.•	١.•
	2 g/L	۲.٦	15.7	1.7	17.7	۲.۷	1.7	۸. •	1.7	٤.•	1.7	٤.•	•.•	• 9
~	4 g/L	Y_V	17	٤.•	۸.۰	1.7	•.•	۰.۳	1.5	۲.٦	•.•	۲.٦	•.•	١.•
Giza	6 g/L	۲.۷	۲.۷	1.5	٤.•	1.5	•.•	1.5	•.•	1.5	•.•	•.•	•.•	•.•
178	8 g/L	•.•	0.7	1.5	۲.۷	•.•	•.•	•.•	•.•	•.•	•.•	•.•	•.•	•_٢
	control	۲۲. ٦	۳٦	١٣.٣	۲٩٫٣	۲.۷	٤.•	٥.٣	٤.•	١٣.٣	۲.۳	٤.•	٤.•	۲. ۱
	2 g/L	۳۱ <u>.</u> ۳	۲٦.٧	٤٠	۳٦	٤.•	٤.•	۳٥	۲.۷	۳٥	•.•	۲٦	1.3	•.•
Sakha	4 g/L	٦٧	۳۱.۳	۲.•	14.5	•.•	٤٠	٦٦	۲٦	٦٦	•.•	1.1	• •	• • •
105	6 g/L	٩٣	١٣٫٣	•.•	٨.•	• •	1.1	٤٠	1.1	٤٠	•.•	•.•	• •	• . ٣
	8 g/L	• .•	٣.٥	•.•	٨. •	•.•	•.•	۳.۲	•.•	•.•	•.•	•.•	• .•	• . ٣
	control	۱۳. ۳	۲۱٫۳	١٠.٧	14.5	٤.٠	٥٣	٨	٤.•	٩٠٣	٦ <sub>.</sub> ٦	٨	٤.•	٠٦
	2 g/L	٦٧	17.0	۲.٧	17.0	۲.٧	٤٠	٦٧	۲.٧	٦٧	۲.٧	٦٦	1.7	• • •
Sakha	4 g/L	٤٠	1.7	۲.٧	٨.•	• •	1.1	۳_٥	•.•	٤٠	•.•	٤٠	• •	• . ٣
106	6 g/L	1.5	٦٧	1.7	۲.٧	•.•	•.•	•.•	•.•	۲٦	•.•	•.•	• .•	• ٢
	8 g/L	• .•	٤.•	•.•	1.1	•.•	•.•	•.•	•.•	۳.۱	•.•	•.•	• .•	• 1
Total Fu	ngi	111	1117	٦0.9	144.9	۲٤	٥.٢٢	٦0.1	۲۱٫۱	۲۱٫٦	١٤.٦	۳۷.۱	۲.۱۰	۷.0
F test		*	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS
L S D 59	%	٥ <sub>.</sub> ٣ ٤	-	-	-	-	-	-	-	-	-	-	-	-

These results agree with earlier studies reported by El-Abbasi, (1990) and Ibrahim, (2003). Aml and Abd El-Hai, (2011) showed that peanut seeds treatment with *S. cerevisiae* inhibited the different seed-borne fungal genera and diseases development under storage conditions up to 6 months. The beneficial effects of yeast extract may be due to the anti fungal activity of its metabolites (Hassanein, *et al.*, 2002), so the application of yeasts as plant pathogens control is recommended where it was found to produce proteinaceous killer lethal to

fungal strains (Hodgson. *et al.*, 1995; Marquina, *et al.*, 2002 and Santos, *et al.*, 2004). Moreover, El-wakil, *et al.*, (2009) stated that disease severity of pre- and post-

emergence damping of caused by *Cephalosporium* sp., *F.* oxysporum, *F.* solani, *R.* solani and *Verticelium dahliae* reduced significantly when seeds of faba bean were coated with water suspension  $(10^9 \text{ cfu mL}^{-1})$  of *S.* cerevisiae before sowing due to activity in disease suppression. Also, El-Tarabily (2004) reported that the fungi activities of *Rhizoctonia solani* diseased sugar beet

plants were well suppressed by using different yeasts. Shalaby, *et al.*, (2008) found that liner growth of *F*. *oxysporum* was inhibited with (39.52%) and (50%) by using 5 g/L and 6.35 g/L of the yeast, respectively. Wide variety of yeasts have been extensively used for the biological control of post-harvest fruits and vegetables diseases (Punja, 1997 and Zheng, *et al.*, 2003), against moulds of stored grains (Petrsson, *et al.*, 1999) and to control powdery mildews (Urquhart and Punja, 1997).

Results in Table 7 illustrated that increase of total fungi lead to decreased in germination percentage and seedling vigor index, Giza 178 cv. seeds total fungi was (92.8), germination percentage (GP) (80%) and seedling vigor index (SVI) (10.4) for the control, while foliar spray with 2, 4, 6, and 8 g/L yeast lead to lower the number of total fungi to (64.7), (40.8), (19.1) and (9.5), respectively and increased (GP) to (90%) and SVI to (18), also on S. 105 cv. seeds was total fungi (140.4), GP (72%) and SVI (17.3) for control and recorded after treatment the less of total fungi with the same concentrations reached to (14.9) for total fungi and increased for GP to (92%) and SVI to (26.7) at 8g/L yeast. On seeds of S. 106 cv. was total fungi (112.4), GP (74) and SVI (13.3) for control, reduction the number of fungi on seeds to (6.7) of total fungi and increased GP to (90%), SVI to (18) with 8 g/L treated. Generally in this table showed that foliar spray with high concentrations of dry yeast increased GP, SVI and reduction numbers of seed-borne fungi associated to rice seeds. These results are similar as of Nagodawithana (1991); Madi *et al.*, (1997); El-Trabily (2004), Shalaby and El-Nady (2008) who reported that *R. solani* and *Sclerotium rolfsii* were effectively suppressed by some plant growth promoting yeasts. Hassan and Abd-Rehim (2002) observed that increasing yeast concentrations (0.05 to 1%) decreased the diseases incidence and controlling onion neck rot disease. *Fusarium* spp. was also inhibited and seed germination of watermelon was induced by several taxa included yeast genera as plant growth promoters and as biocontrol agents (Lokesh, *et al.*, 2007).

# CONCLUSION

It could be concluded that use of foliar spray with dry yeast(*S. cervisia*) at 6 - 8 g/L at 25, 55 and 85 days after rice sowing gave the highest values of yield, yield components and next enhanced seed, seedling vigor and lower numbers of seed-borne fungi associated with rice seeds (seed quality).

 Table 7 : Effect of foliar spray with yeast concentrations on the total fungi, germination percentage ,and seedling vigor index of three rice cultivars.

Yeast Concentrations		Giza 17	8		Sakha 1	05	Sakha 106			
	Total Germination		Seedling vigor	Total	Germination	Seedling vigor	Total	Germination	Seedling vigor	
	fungi	(%)	index (SVI)	fungi	(%)	index (SVI)	fungi	(%)	index (SVI)	
Control	٩٢٨	۸.	۱۰.٤	١٤٠ ٤	77	14.5	117.5	٧٤	۳.۳	
2 g/L	٦٤.٧	71	15.1	175.7	۸.	17.4	٧٥.٣	٨٢	17.5	
4 g/L	٤٠٨	٨٤	17.0	٦٨٩	٨٤	77.7	٤٠٢	71	١٤.٨	
6 g/L	19.1	$\wedge \wedge$	17.1	٤١.٥	٨٤	۲۱.۰	15.1	71	11.9	
8 g/L	٩٫٥	٩٠	۱۸.۰	١٤.٩	٩٢	۲٦.٧	٦.٧	۹.	۱۸.	

# REFERENCES

- Abdel-Hakim, W. M.; Y. M. M. Moustafa and R. H. M. Gheeth (2012). Foliar application of some chemical treatments and planting date affecting Snap Bean (*Phaseolus vulgars* L.) plants growth in Egypt. J. of Hort. Sci. & Ornamental Plants, 4 (3): 307-317.
- Agarwal, P. C.; C. M. Mortensen and S. B. Mathur (1989). Seed –borne Diseases and Seed Health Testing of Rice Tech. Bulletin No. 3, Phytopathological Paper No. 30 CAB International Mycological Institute (CMI) Kew, Surrey,UK., pp :58-59.
- Agarwal, V. K. & A. K. Srivastava (1985). NaOH seed soak method for routine examination of rice seed lots for rice bunt. Seed Res., 13 : 159.
- Agrawal, V. K. and A. K. Srivastava (1981). A simpler technique for routine examination of rice seed lots for rice bunt. Seed Technology News, 11: 1-2. Bull. of the Indian Soci. of Seed Tech.
- Amer, S. S. A. (2004). Growth, green pods yield and seeds yield of common bean (*Phaseolus vulgaris* L.) as affected by active dry yeast, salicylic acid and their interaction. J. Agric. Sci., Mansoura Univ., 29 (3): 1407-1422.

- Aml, E. A. El-Saidy and K. M. Abd El-Hai (2011). Alleviation of Peanut seed deterioration during storage using Biotic and Abiotic agents. Res. J. of Seed Sci., 4 (2): 69-81.
- AOSA., (1983). Seed Vigor Hand Testing Book. Contribution No. 32 to the Handbook on Seed Testing. Association of Official Seed Analysis, Springfield, IL
- Barnett, J. A.; R. W. Payne and D. Yarrow (1990). Yeasts characteristics and identification. Cambradge . Camb. CBZBR, pp 999.
- Booth, C., (1985). The Genus *Fusarium* Commonwealth Mycological Institute, Kew. Surrey, England,: 237pp.
- Burrges, L. W.; C. M. Liddell and B. A. Summerell (1988). Incorporating a key and descriptions of common species found in Australasia (second edition) *Fusarium* Rese. Lab. Depart. of Plant Path. and Agric. Entomo. Univ. of Sydeny, PP: 156.
- Chidambaram, P.; S. B. Mathur and P. Neergaard, (1973). Identification of seed-borne *Drechslera* sp. Danish Government Institute of seed patho. for Developing Countries, Hellerup, Copenhagen Denmark, pp: 207.

- Cook, R. J. (1993). Making greater use of introduced microorganisms for biological control of plant pathogens. Annu. Rev. Phytopathol., 31: 53-80.
- Darweesh, M. A.; E. A. Tartoura and K. Dawa (2003). Effect of phosphorous fertilization and some growth promoters on growth and yield of pea. J. Agric. Sci. Mansoura Univ. Egypt, 28 (12): 1327-1343.
- El-Abbasi, I. H. (1990). Studies on some seed borne disease of certain food crop in Egypt. M. Sc. Thesis, Fac. Agric., Ain Shams Univ., 83 pp.
- El-Bassiony, A. M.; Z. F. Fawzy; M. A. El-Nemr and Li Yunsheng (2014). Improvement of growth, yield and quality of two varieties of Kohlrabi plants as affected by application of some bio-stimulants. Middle East J. of Agric., Res., 3(3): 491-498.
- El-Emery, G. A. E. (2004). Effect of growth regulators of yeast autolysate, RNA and adenine on some seeds during germination. Arab Uni. J. of Agric. Sci., 12(1): 51-67.
- El-Ghamriny, E. A.; H. M. H. Arisha and K. A. Nour (1999). Studies in tomato flowering, fruit set, yield and quality in summer season, 1. Spraying with thiamine, ascorbic acid and yeast. Zagazig, J. Agric. Res., 26: 1345-1364.
- Ellis, M. B. (1980). Dematiaceous Hyphomycetes. Commonwealth Mycological Institute, Kew, Surry, England.
- El-Sirafy, Z. M.; A. M. Abd El-Hameed and Rasha E. H. El-Mahdy (2011). Role of biofertilization and different rates of nitrogen with foliar spraying nitrogen and yeast in rice productivity and nutrient uptakes. J. Soil Sci. and Agric. Eng., Mansoura Univ., 2 (7): 717-731.
- El-Tarabily, K. A. (2004). Suppression of *Rhizoctonia* solani disease of sugar beet antagonistic and plant growth promoting yeasts . J. Appl. Microbiol., 96 : 69-75.
- Elwakil, M. A.; O. A. Awadallah; I. M. El-Refai; M. A. El-Metwally and M. S. Mohammed (2009). The use of bread yeast as a biocontrol agent for controlling seed-borne fungi of faba bean. Plant Pathol. J., 8: 133-143.
- Entian, K. D and K. U. Fröhlich (1984). Saccharomyces cerevisiae mutants provide evidence of hexokinase P11 as a biofunctional enzyme with catalytic and regulatory domains for triggering carbon catabolite repression. J. Bacteriol., 158 (1): 29-35.
- Fakir, G. A.; I. Hossain; M. U. Ahmed; M. A. U. Doula and M. M. Alam (2002). Quality of farmer's boro and T. aman rice seeds collected before sowing from Bogra, Ragshahi and Rangpur district of Bangladesh. Proc. of the 2002 Planning Meeting of the Rice seed health Improvement Sup-Project, April 17 - 18, 2002, Gazipur, Bangladesh.
- Fathy, E. S. L. and S. Farid (1996). The possibility of using vitamin B and Yeast to delay senescence and improve growth and yield of common beans (*Phaseolus vulgaris* L.). J. Agric. Sci. Mansoura Univ., 21 (4): 1415-1423.
- Ghoname, A. A.; M. A. EL-Nemr; A. M. R. Abdel-Mawgoud and W. A. El-Tohamy (2010).

Enhancement of sweet pepper crop growth and production by application of biological, organic and nutritional solutions. Res. J. of Agric. and Biol. Sci., 6 (3): 349-355.

- Glick, B. R. (1995). The enhancement of plant growth by free living bacteria. Cand. J. Microbio., 41: 109-117.
- Gomez, K. A. and A. A. Gomez (1984). Statical procedures for agricultural research 2nd Ed. John Waliy and Sons.
- Guerrero, F. C.; S. B. Mathur and P. Neergaard (1972).
  Seed Health Testing of Rice: Seedborne fungi Associated with Abnormal Seedling of Rice.Vol. 37, International Seed Testing Association, Switzerland, pp: 985-997.
- Hassan, M. H. A. and G. H. Abd El-Rehim (2002). Yeast application as a biofertilizer and biocontrol agent for Onion neck rot disease in relation to bulb productivity and quality. Assiut Agric. Sci., 33 (1): 241-251.
- Hassanein, M. M.; A. A. El-Mehalawy; H. M. Khater; A. K. El-Zahraa and A. Y. Youssef (2002). The potential of selected Rhizosphere actinomycetes and yeast fungi for the biological control of late wilt disease of maize caused by *Cephalosporium maydis*. Afircan. J. Mycol. Biotech., 1:167-188.
- Hodgson, V. J.; D. Button and G. M. Walker (1995). Anti-candida activity of a novel killer toxin from the yeast (*Williopsis mraki*). Microbiology, 141: 2003-2012.
- Ibrahim, E. A. M. (2003). Studies on seed-borne diseases fungi of rice and its control. M.Sc. Thesis, Fac. Agric., Al-Azhar Univ., 102 pp.
- ISTA (1999). International Rules for Seed Testing Handbook of Vigor Test Method. 3rd Edition: 22-25.
- Khedr, Z. M. A. and S. Farid (2000). Response of Naturally virus infected tomato plants to yeast extract and phosphoric acid application. Annals of Agric. Sci., Moshtohor, 38 (2): 927-939.
- Lokesh, S.; B. G. Bharath; V. B. Raghavendra and M. Govindappa (2007). Importance of plant growth promoting rhizobacteria in enhancing the seed germination and growth of watermelon attacked by fungal pathogens. Acta Agronomica Hungariea, 55(2): 243-249.
- Madi, L.; T. Katan ; J. Katan and Y. Heins (1997). Biological control of *Sclerotium rolfsii* and *Verticillium dahaliae* by *Talaromyces flaves* is mediated by different mechanisms. Phytopatho., 87 (10):1054-1060.
- Mady, M. A. (2009). Effect of foliar application with yeast extract and zinc on fruit setting and yield of faba bean (*Vicia faba*). J. Biol. Chem. Environ. Sci., 4 (2): 109 127.
- Mahmoud, T. R. (2001). Botanical studies on growth and germination of Magnolia (*Magnolia grandifbra* L.) plants. M.Sc. Thesis, Fac. Agric. Moshtohor, Zagazig Univ., 103 pp.
- Marquina, D.; A. Santos and J. M. Peinado (2002). Biology of killer yeast. Int. Microbiol., 5: 65-71.

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- Mohamed, A. N.; E. U. Mohamed and A. N. Samah (2015). Influence of foliar spray with yeast extract on vegetative growth yield of fresh herb, anatomical structure, composition of volatile oil and seed yield components of Basil plant (*Ocimum basilicum* L.). Intern. J. of Adv. Res., 3(10): 978-993.
- Muller, K. and A. C. Leopold (1966). Correlative aging and transport of p 32 in corn leaves under the influence of Kinetin Plant, 68: 167-185.
- Nagodawithana, W. T. (1991). Yeast technology. Universal foods cooperation Milwauke, Wisconsin. Published by Van Nostrand, New York.
- Natio, K. ; S. Nagamo ; K. Fury and H. Suzki (1981). Effect of benzyladinine on RNA and protein synthesis in intact bean leaves at various stages of ageing. Plant Physiol., 52: 342-348.
- Petersson, S.; N. Jonsson and J. Schnurer (1999). *Pichia* anomata as a biocontrol agent during storage of high-moisture feed grain under conditions. Postharvest Biol. Technol., 15.
- Punja, Z. k. (1997). Comparative efficacy of bacteria, fungi and yeasts as biological control agents for disease of vegetable crops. Can. J. Plant Pathol., 19: 315-323.
- Ram, S.; S. D. Dodan and P. O. Sheoran (1998). Kernel smut of rice: present status. Intern. J. of Tropical Plant Dis., 16 (2): 149-167.
- Rania, M. A. Nassar; Y. M. Ahmed and D. M. A. Nassar (2011). Effect of foliar spray with active yeast extract on morphological, anatomical and yield

characteristics of Kidney Bean (*Phaseolus vulgaris* L.). Australian J. of Basic and Appl. Sci., 5(5): 1071-1079.

- Reyes, G. M. (1933). The black smut or bunt of rice (*Oryza sativa* L.) in the Philippines. Philippines J. of Agric., 4: 241-270.
- Said, A. S.; F. F. Zakaria and R. E. Hassan (2012). Response of cucumber plants to foliar application of chitosan and yeast under greenhouse conditions. Australian J. of Basic and App. Sci., 6(4): 63-71.
- Santos, A.; A. Sanchez and D. Marquina (2004). Yeasts as biological agents to control *Botrytis cinerea*. Microbiol. Res., 159: 331-338.
- Shalaby, M. E. and M. F. El-Nady (2008). Application of Saccharomyces cerevisiae as a biocontrol agent against Fusarium infection of sugar beat plants. Acta Biologica Szegediensts, 52(2): 271-275.
- Sharma, R. C. and H. S. Chahal (1996). Seed health studies on parental lines of hybrid rice. Seed Res., 24(2) 145-150.
- Tarp, G.; L. Lange and O. Kongsdol (1987). Seed-borne pathogens of major food crops in Mozambique, Seed Sci. and Technol., 15: 793-310.
- Urquhart, E. G.; Z. K. Punja (1997). Epiphytic growth and survival of *Tilletiopsis pallescens*, a potential biological control agent of *Sphaerotheca futiginea*, on cucumber leaves .Can. J. Bot., 75 : 892-901.
- Zheng, D. X.; Y. P. Sun and H. Y. Zhang (2003). Yeast application for controlling apple post harvest diseases associated with *Penicillium expansum*. Bot. Bull. Acad. Sin., 44: 211-216.

تأثير الرش الورقي بالخميرة على محصول وجودة تقاوي بعض أصناف الأرز جلال احمد دويدار \* و السيد احمد محمد إبراهيم\*\* \* قسم بحوث الأرز - معهد بحوث المحاصيل الحقلية - مركز البحوث الزراعية - مصر \*\* قسم بحوث تكنولوجيا البذور – معهد بحوث المحاصيل الحقلية - مركز البحوث الزراعية - مصر

أجريت تجربتي حقل ومعمل لدراسة تأثير الرش الورقي بالخميرة الجافة بتركيزات صفر ٢ ٢ ٤ ٢ ٢ ٨ جم/ لتر في أعمار ٢٠ ٥٩، ٥٠ يوم من الزراعة على نمو المحصول ومكّوناته وجودة التقاوي المنتجة من أصناف الأرز جيزة ١٧٨ وسخا ١٠٥ وسُخاً ١٠٦ لمُوسمين ٢٠١٢ – ٢٠١٣ وذلك بمُحطة بحُوث السرو– مركز البحوث الزراعية أجريت التجربة ألحقليه في أربع مكررات بتصميم شرائح متعامدة والتجربة ألمعمليه لاختبارات جودة التقاوي على ناتج التجربة ألحقليه بتصميم التام العشوائية في أربع مكررات بوحدة بحوث تكنولوجيًّا البذور بالمنصورة – معهد بحوث المحاصيل ألحقليه – مركز البحوث الزراعية وأدت النتائج إلى الآتي 🗧 -أظهرت النتائج بالرش الورقي بالخميرة خاصة التركيرات الأعلى ٦ , ٨ جم/لتر إلى زيادة معنوية على العديد من صفات النمو والمحصول وخاصة مع صنف جيزة ١٧٨ مثل عدد الأفرع /٢ وطول النبات (سم) والوزن الجاف (جم/ ٢٦) وعدد السنابل / م ٢ وطول النبات (سم) وعدد الحبوب الممتلئة /السنبلة ووزن السنبلة (جم) ومحصول الحبوب (طن/هكتار) ومحصول ألقش (طن/هكتار) يليه صنف سخا ١٠٦ ثم صنف سخا ١٠٥ كمقارنة أصناف ٢٠- أدى الرش الورقي بالخميرة بزُيادة التركيزات من صفر إلى ٢ ٤ ، ٢ ، ٨ جم/ لتر إلى زيادة وتحسن في صفات مكونات المحصول مثل عدد الأفرع /٢ م وأيضا تاريخ الطرد (يوم) وزيادة الوزن الجاف (جم/ م٢) و عدد السنابل / م٢ وطول النبات (سم) ووزن الألف حبه بالجرام ومحصول الحبوب (طن/هكتار) بفروق معنوية عاليه مقارنة بالكنترول لكلا الموسمين وأيضا كَانَ التفاعل معنوي مع صفات عدد الأفرع / م٢ وعدد السنابل / م٢ ووزن الألف حبه بالجرام ومحصول الحبوب (طن/هكتار) مع كل الأصناف تحت الدراسة . ٣-اختبار حبوية التقاوي :- حيث كان صنف جيزة ١٧٨ هو أعلى الأصناف لصفات البادرات الطبيعية (نسبة الإنبات) وسرعة الإنبات وطول البادرات بينما يليه صنف سخا ١٠٩ ثم سخا ١٠٦ كمقارنة أصناف ومن ناحية أخرى كان استخدام تركيز ٦ و ٨ جم/لتر خميرة كرش ورقى أعطى تحسن معنوي لحيوية البذور و البادرات وأيضا أدى إلى خفض معنوي للنسبة المنوية للبادرات الغير طبيعيه. ٤- اختبار صحة التفاوي :- تم تعريف عدد ١٣ جنس ونوع فطرًى تم ملازمتهم لتفاوي كل الأصناف تحت الدراسة وسجَّل صنف جيزة ١٧٨ اقل الأصناف حملا للفطريات يليه صنف سَّخا ١٠٦ ثم سخا ١٠٥ كمقارنة أصنّاف ومن ناحية أخرى كان فطر Alternaria longesima أكثر الفطريات تواجدا على البذور يليه فطر Bipolaris oryza ثم Alternaria longesima ثم Alternaria padwickii Sarocladium oryza ,F. graminearum ,tenuis, وأدت الزيادة في تركيز الخميرة كرش ورقى من صفر إلى ٢, ٤, ٦, ٨ جم/ لتر إلى نقص معنوي في أعداد الفُطريات ألملازمه للتقاوي وخاصة الممرض منها والمسبب للنقص في حيوية البذور والبادرات . التوصية :-

ي توصى هذه الدراسة باستخدام الخميرة الجافة كرش ورقى على نباتات الأرز في أعمار ٢٥, ٥٥, ٨٥ يوم من ألزراعه بتركيز ٦ – ٨ جم/لتر حيث يؤدى ذلك إلى تحسن في صفات النمو والمحصول ومكوناته وارتفاع في قيم حيوية البذور والبادرات ونقص في أعداد الفطريات المصاحبة للتقاوي ألمنتجه .