


#### Abstract

Intraspecific hybridization among four cultivars of snapdragon, i.e. $P_{1}=$ Sonnet wit (white), $\mathrm{P}_{2}=$ Sonnet karmijn (red), $\mathrm{P}_{3}=$ Sonnet rose (rose) and $\mathrm{P}_{4}=$ Sonnet geel (yellow) was carried out during three successive growing seasons: 95/1996, 96/1997 and 97/1998 at Antoniadis Botanical Garden, Horticulture Research Institute, Agriculture Research Center, Alexandria, Egypt.

The main objective of this investigation was to produce new patterns of snapdragon, especially with reference to flower characteristics. Diallel cross analysis was used to study and determine the genetic system controlling vegetative growth and also the type of gene action for the different traits.

Additive and dominance gene effects were significant in the $F_{1}$ and $F_{2}-$ generations with respect to plant height. The overdominance gene effects played an important role in the inheritance of plant height. Heritability in narrow sense was intermediate in both generations and no inbreeding depression was obtained for most crosses. Most of dominant genes have positive effects and increased plant height. All $F_{1}$ - crosses achieved positive heterosis.

Additive gene effect played the major role by the inheritance of the number of branches per plant. Heritability in narrow sense was intermediate in the $F_{1}$ and $F_{2}$ generations. Most of dominant genes have negative effects and decreased the number of branches. The overdominance gene effects played an important role for $\mathrm{F}_{1}$ hybrids and partial dominance in the $F_{2}$ generations. The dominant genes were more frequent than recessive in the parents. Most crosses achieved positive heterosis.

Additive and dominance genes effects were involved in the inheritance of the number of leaves per plant. Heritability in narrow sense was moderate indicating that this trait could be advanced by selection. The degree of dominance $\left(H_{1} / D\right)^{1 / 2}$ was found to be overdominance in the $\mathrm{F}_{1}$ hybrids confirmed with $\mathrm{Wr}, \mathrm{Vr}$ graph. In the $\mathrm{F}_{2}$ generations the results of Wr , Vr graph contradicted with the ratio $\left(\mathrm{H}_{1} / \mathrm{D}\right)^{1 / 2}$ indicated the presence of epistasis. Dominance genes seemed to be acted in negative direction in the $F_{1}$ hybrids and $P_{1}$ carried most recessive genes, while in the $F_{2}$ dominance genes acting in positive direction and $\mathrm{P}_{2}$ and $\mathrm{P}_{3}$ have most dominant genes.

In the $F_{1}$ hybrids, additive and dominance components were found to be not significant with respect to leaf area. Environmental effects were high and such large environmental role and also overdominance was involved in the inheritance of leaf area. The parent seemed to carry more dominant genes than recessive. Dominant genes seemed to be acting in positive direction and increased leaf area.


## INTRODUCTION

Snapdragon (Antirrhinum majus) is widely cultivated in gardens and houses. It is used as a cut flower, pot plant and for cultivation in flowerbeds. The flowers of snapdragon have an unfamiliar and attractive shape and colours. The flowers are arranged in simple racemes or spikes. The plant

Badr, M. et al.
may be tall or short, upright or semi spherical according to varieties. (ElGamassy and Nada, 1974).

The genetics and inheritance of plant height, the number of branches per plant, the number of leaves per plant as well as leaf area have been biometrically analyzed and studied in Antirrhinum majus (El-Torky, 1981 and Misiha, 1991) as well as in many other plants (Ahmed and Ismail, 1999; Deore et. al., 1997 and Kumar et al., 1998).

Estimation of genetic variances and its components are the first step to determine the most appropriate breeding scheme, the choice of an efficient breeding method on the magnitude and type of gene effect prevailing in the population under study. The diallel cross analysis of Hayman (1954 and 1957) is a useful method to divide the phenotypic variation into genotypic and environmental components and further subdivide the genotypic variation into additive and non-additive components. To improve any quantitative character which show a continuous range of variation, information about the gene action of the character has to be acquired. The different gene actions involved in the inheritance of such characters are additive (resulting from average effects of genes) and non - additive (resulting from dominance and epistasis effects among the genes). The relative importance of these two components provides the breeder with valuable information about the possibilities and methods of improving these characters. If the additive gene action appears to be more important contributor to the genetic variability of a character, a maximum improvement in this particular character must be expected by the breeder through a carefully designed selection programme. On the contrary, the presence of a relatively high non - additive gene suggests that a hybrid programme will perform good prospects for the characters under consideration, as a result of a direct relationship between the non - additive gene action and heterosis (Jinks, 1954).

## MATERIALS AND METHODS

The effects of crossing different cultivars of snapdragon (Antirrhinum majus, L.) on the characteristics of vegetative growth were studied throughout three generations, i.e. parental, first and second generations. The experiments were carried out during three successive growing seasons of 95/1996, 96/1997 and 97/1998 at Antoniadis Botanical Garden, Horticulture Research Institute, Agriculture Research Center, Alexandria, Egypt.

Certified seeds of four snapdragon cultivars; $P_{1}=$ Sonnet wit (white), $P_{2}=$ Sonnet karmijn (red), $P_{3}=$ Sonnet rose (rose) and $P_{4}=$ Sonnet geel (yellow) were obtained from Hamer Bloemzaden b.v., Holland.

Seeds of parental cultivars (first season) were sown on December 10, 1995. Seedlings were transplanted on March 5, 1996. As soon as the plants started to flower, all possible crossing combinations were made to obtain the $F_{1}$ - seeds. The $F_{1}$ - seeds were sown on December 8,1996 . The $F_{1}$ - young plants were transplanted on February 25, 1997. As soon as the $F_{1}$ - plants started to bloom, selfings were carried out to obtain the $F_{2}$ - seeds which were sown on October 18, 1997 and transplanted on December 10, 1997.

The layout of the experiments was a randomized complete block design with three replications (Steel and Torrie, 1986). Each replication contained 16 selfings and crosses (16 genotypes) and every selfing and cross consisted of 36 plants.

The collected data included :

1. Plant height (in cm .) measured from the soil surface to the top of the longest branch.
2. Number of branches per plant.
3. Number of leaves per plant.
4. The leaf area (in $\mathrm{cm} .^{2}$ ) expressed as the average mean weight of a leaf divided by the mean weight of one $\mathrm{cm} .{ }^{2}$.

The nature and the amount of genetic parameters were performed by Hayman's approach (Hyman, 1954 and 1957), which was used to divide phenotypic variation into genotypic and environmental components. The detailed description of the various genetic properties and parameters were calculated after Singh and Chaudri (1977).

## RESULTS AND DISCUSSION

## 1. Plant height

Statistical analyses proved that $P_{3}$ was the tallest among the parents and it differed significantly from all parents (Table 1). Crosses derived from the $P_{3}$ in either the parental or the maternal directions were taller than the parents in both the first and second seasons except the cross $P_{3 \times} P_{2}$. All crosses exhibited significant positive heterosis estimates (Table 2). There were no differences between the $\mathrm{F}_{1}$-crosses and their reciprocals except those of the $P_{1} \times P_{4}$ and $P_{4} \times P_{1}$. In the $F_{2}$ generations, most of the crosses and their reciprocals showed no differences with two exceptions, i.e. $\mathrm{P}_{1} \times \mathrm{P}_{4}$ and $P_{3 \times} \times P_{4}$ and their reciprocals. The difference was due to the maternal effect. Most crosses gave no inbreeding depression as shown in Table 2 except for $\mathrm{P}_{3 \times} \times \mathrm{P}_{2}$ and $\mathrm{P}_{2 \times} \times \mathrm{P}_{4}$ indicating that additive gene effect was important for plant height and also dominance gene effect in some crosses as reported also by Mahdy et al. (1983) on cotton and Abdel- Sabour et al., (1996) on wheat.

The assumptions of Jinks (1954) and Hayman (1954) of diploid segregation and homozygous parents were found valid and no reciprocal differences may be considered valid with some degrees of confidence. The method of testing the assumptions of no epistasis, no multiple alleles and uncorrelated gene distribution, was done through the regression coefficient (b) by calculating the regression of covariance on the variance; b was found to be significantly different from zero ( $\mathrm{b}=0.97 \pm 0.09$ in the $\mathrm{F}_{1}$ and $0.92 \pm$ 0.19 in the $F_{2}$ ) and is not significantly different than 1.0 (Fig. 1), therefore the assumptions were valid as reported by Misiha (1991) on Antirrhinum majus.

Genetic parameters presented in Table 3 indicated that the dominance gene effect " $\mathrm{H}_{1}$ " as well as the additive gene effect " D " were significant in both $F_{1}$ and $F_{2}$ indicating their importance in the inheritance of plant height. On the other side, " $E$ " component estimating environmental

Badr, M. et al.
$1+2$
J. Agric. Sci. Mansoura Univ., 25 (10), October, 2000. 3

Badr, M. et al.

Fig1+2
effect was significant only in the $\mathrm{F}_{1}$ indicating minor effect of environment on plant height variation. These results agreed with the finding of Misiha (1991) on Antirrhinum majus and Madic (1996), on barley. The proportion $\left(\mathrm{H}_{2} / 4 \mathrm{H}_{1}\right)$ was 0.25 in the $F_{1}$ and KD/KR preparation was found to be close to the unity indicating that the dominant and recessive genes were distributed in equal proportions among the parents, while In the $\mathrm{F}_{2}, \mathrm{H}_{2} / 4 \mathrm{H}_{1}$ was less than 0.25 indicating a symmetry of positive and negative gene proportions in the parents with KD/KR which was larger than the unity indicating that the parent carry more dominant than recessive genes.
$\left(\mathrm{H}_{1} / \mathrm{D}\right)^{1 / 2}$ estimate of the degree of dominance suggested overdominance in both generations. This result is supported by the finding shown in Wr , Vr graph (Fig1), where the regression line intercepted the Wr axis in a negative position in both $F_{1}$ and $F_{2}$ generations which agreed with results of Bakheit and Ezzat (1987) on sesame. In the $F_{1}, ~ " P_{3}$ " had most dominant genes and $P_{1}$ and $P_{4}$ had most recessive genes, while $P_{2}$ had equal frequency of dominant and recessive genes. In the $F_{2}, P_{3}$ and $P_{2}$ had most dominant genes, while $P_{4}$ had most recessive genes and $P_{1}$ had equal frequency of dominant and recessive genes. Correlation coefficient of $\mathrm{Wr}+$ Vr and Vr was negative suggesting that most of dominant genes had positive effect and increased plant height, which agreed with the results of Weber (1976) on peas and El-Torky (1981) on Antirrhinum majus.

The estimates of heritability in broad sense were high (Table 3) indicating that plant height was a genetically controlled character, while the narrow sense heritability estimates were moderate in both generations reflecting the moderate magnitude of additive gene effect in the inheritance of plant height.

## 2. Number of branches per plant

The mean values for the number of branches per plant are presented in Table1. The data showed that $P_{1}$ and $P_{2}$ achieved the highest number of branches in the $F_{1}$ and $F_{2}$ and they were statistically different from $P_{3}$ and $P_{4}$, which had the lowest number of branches. By the $\mathrm{F}_{1}$-progenies, $\mathrm{P}_{2} \times \mathrm{P}_{4}$ and $P_{3} \times P_{4}$ and their reciprocals as well as $P_{3} \times P_{2}$ and $P_{2} \times P_{1}$ produced more branches than their parents, they had also positive heterosis values expressing a clear hybrid vigour (Table 2), while $P_{3} \times P_{1}, P_{1} \times P_{4}$ and $P_{4 \times} P_{1}$ were similar to their parents producing comparatively low number of branches and had also negative heterosis values. The $\mathrm{F}_{2}$-progenies of all crosses were intermediate between their parents except those of $P_{1 \times} P_{2}, P_{3} \times P_{4}, P_{3} \times P_{4}$ and their reciprocals which had higher number of branches compared to their parents.

Inbreeding depression (I.D.) estimates (Table 2) were found to be positive for each of the crosses $\mathrm{P}_{1} \times \mathrm{P}_{3}, \mathrm{P}_{2} \times \mathrm{P}_{4}, \mathrm{P}_{3} \times \mathrm{P}_{4}$ and their reciprocals as well as for the $P_{3} \times P_{2}$ progeny. The other crosses achieved negative I.D. estimates indicating that additive gene effect had a major effect in the inheritance of the number of branches per plant and that the dominant gene effect was also involved.

Due to the presence of significant differences between crosses, the diallel analysis was employed to the data according to the method of Hayman

## Badr, M. et al.

(1954) to test the validity of the assumption of analysis. It could be concluded that diploid segregation, homozygous parents and no reciprocal differences have been already proven to be valid. The assumptions, no epistasis, no multiple alleles and uncorrelated gene distribution could be tested by calculating (b) which was not found to differ significantly from 1.0 ( $b=0.96+$ 0.36 ), while in the $F_{2} b=0.81+0.02$ indicating that these assumptions were not fulfilled (Fig. 2). Estimates of genetic parameters and ratios were calculated and presented in Table 3 . The " $\mathrm{H}_{1}$ " component estimating dominance was found to be significant in the $F_{1}$ and non-significant in the $F_{2}$, while " D " component was significant in the two seasons indicating that the additive gene effect played a major role in the inheritance of this trait, which agreed completely with the findings of Misiha (1991) on Antirrhinum majus and Cinsoy (1992) on soybean. The "E" component was not significant either in $F_{1}$ or in $F_{2}$ indicating that there was minor effect of environment in the variation of this trait. There was a symmetry of positive and negative gene proportion in the parents from ratio $\mathrm{H}_{2} / 4 \mathrm{H}_{1}$ supported by KD/KR, which was larger than 1.0 in the $F_{1}$ - generation indicating that parents carry more dominant genes than recessive, while in the $F_{2}$, there was equal distribution in the parents according to $\mathrm{H}_{2} / 4 \mathrm{H}$ which was larger than 0.25 and confirmed also by KD/KR, which was equal to the unity. With regard to $\left(H_{1} / D\right)^{1 / 2}$, estimation was close to 1.0 in Findicating the presence of overdominance, while in the $F_{2}$, it was found to be lower than 1.0 suggesting a partial dominance. These results agreed with the ( $\mathrm{W}_{\mathrm{r}}, \mathrm{V}_{\mathrm{r}}$ ) graph as shown in Fig 2. The regression line intercepted the $W_{r}$ axis in a position near to the origin expressing a complete dominance case in $F_{1}$, while in $F_{2}$, the regression line intercepted Wr axis in positive position indicating a partial dominance. The Wr, Vr points corresponding to the parents $P_{3}$ and $P_{4}$ fall near to the point of origin suggesting that these parents carried most dominant genes, while $P_{1}$ carried most recessive genes and $P_{2}$ had nearly equal frequency of dominant and recessive genes in both $F_{1}$ and $F_{2}$. Moreover, the $(\mathrm{Wr}+\mathrm{Vr})$ values were positively correlated with the parental means indicating that most of dominant genes had negative effect and decreased the number of branches per plant. With regard to $\mathrm{h}^{2} / \mathrm{H}_{2}$ ratio, it indicated that one group of gene exhibiting dominance was found to control the number of branches.

Heritability in broad sense was high in the $F_{1}$ and $F_{2}$ generations (Table 3) indicating that this character was genetically controlled. In the same time heritability in narrow sense was moderate in $F_{1}$ and $F_{2}$ referring to the presence of additive gene effect, so this character could be advanced by selection as reported by Yadav and Chankar (1991) on okra and Misiha (1991) on Antirrhinum majus.

## 3. Number of leaves per plant

The mean values of the number of leaves per plant presented in Table1 indicated that the parents differed greatly from each other. The parent $\left(\mathrm{P}_{1}\right)$ produced the highest values for $\mathrm{F}_{1}$ and $\mathrm{F}_{2}$ followed by $\mathrm{P}_{2}$.

The crosses which involved $P_{2}$ always gave high values in the $F_{1}$-and $F_{2}$ - generations and achieved positive heterosis values (Table 2), with one
exception ( $\mathrm{P}_{2} \mathrm{XP} \mathrm{P}_{1}$ ), whereas $\mathrm{P}_{1}$ gave only one case of positive heterosis by its cross with $P_{2}$, all other crosses achieved negative values.

There were no differences between the crosses and their reciprocals except those of $P_{1} \times P_{3}$ and $P_{3} \times P_{4}$ in the $F_{1}, P_{1} \times P_{2}$ and $P_{1} \times P_{4}$ in the $F_{2}$.

The importance of additive gene effects on the number of leaves per plant was obviously detected from Table 2 due to the negative values obtained for inbreeding depression (I.D) in most cases. Only two crosses gave positive values, i.e., $P_{2} \times P_{3}$ and $P_{3} \times P_{4}$, which gives an indication to the possible role of dominance genes as supported by Singh and Sudhir (1996) on Papaver somniferum.

As reviewed previously, diallel cross analysis suggested by Hayman (1954) was based on several assumptions. No epistasis, no multiple allele, and uncorrelated gene distribution are valid. These results confirmed with the regression coefficient of Wr upon Vr which did not differ significantly from 1.0 in $\mathrm{F}_{1}$ and $\mathrm{F} 2(\mathrm{~b}=1.02 \pm 0.26,1.03 \pm 0.14$ respectively) and it differed significantly from Zero in $\mathrm{F}_{2}$ (Fig. 3). The other assumptions of diploid segregation, homozygous parents were valid, no reciprocal differences assumption was valid with some degrees of confidence. The results presented in Table 3 showed that the additive genetic component " $D$ " was significant in $F_{1}$ and $F_{2}$ and also dominance genetic component " $\mathrm{H}_{1}$ " was significant indicating the importance of additive gene and dominance gene for this character. These results agreed with Shamsuddin et al. (1980) on tobacco. The "E" component estimating the environment effect was significant in $\mathrm{F}_{2}$ only. The $\mathrm{H}^{2} / 4 \mathrm{H}_{1}$ ratio was less than 0.25 in both generations, indicating a symmetry of positive and negative genes in the parents and that the parents carry more dominant genes than recessive reflecting proportion KD/KR, which was larger than one in F1 but the parents seemed to carry more recessive genes than dominant in $F_{2}$, where KD/KR was found to be less than 1.0. The proportion $\left(H_{1} / D\right)^{1 / 2}$ in $\mathrm{F}_{1}$ was larger than 1.0 indicating overdominance. This could be confirmed with the $\mathrm{Wr}, \mathrm{Vr}$ graph as shown in Fig. 3 where the regression line intercepted $W r$ axis in negative position, while in $F_{2}\left(H_{1} / D\right)^{1 / 2}$ was less than 1.0 indicating partial dominance which disagreed with Wr , Vr graph, where regression line intercepted Wr axis in negative position indicating overdominance, since epistasis can decrease or increase the average degree of dominance (Hayman, 1957). In addition, Mather and Jinks (1971) reported that the ratio $\mathrm{H}_{1} / \mathrm{D}$ is not a measure of degree of dominance, so this estimate is not true and the graphic analysis may be reliable. The $\mathrm{Wr}, \mathrm{Vr}$ points corresponding to the parents in the $F_{1}$ for $P_{4}$ and $P_{3}$ had most dominant genes, while $P_{1}$ had more recessive genes, but in $F_{2}, P_{4}$ carried most recessive genes, while $P_{1}, P_{3}, P_{2}$ where found to be in intermediate position.

Correlation coefficient between parental means and $\mathrm{Wr}+\mathrm{Vr}$ was positive in $F_{1}$ indicating that most of the genes exhibiting dominance were associated with the lower number of leaves, while in $F_{2}$, the position was reversed.

Heritability in broad sense was high (Table 3). This finding indicated that the number of leaves per plant is under the control of genes, while heritability in narrow sense was found to be moderate in the $F_{1}$ and $F_{2}$, which

## Badr, M. et al.

agreed with the results of Rastogi et al. (1995) on Chinese cabbage and Khar et al. (1997) on cauliflower.

## 4. Leaf area

Parents were not significantly different from each other, while in the $F_{2}$ generations, $P_{3}$ produced the highest value (Table 1). Progenies of all crosses achieved higher values for leaf area compared to their parents. They also obtained positive heterosis values indicating the presence of hybrid vigour for all crosses (Table 2). No significant differences were found between crosses and their reciprocals except in the case of $P_{1} \times P_{3}$ in the $F_{1}$ and $P_{1} \times P_{3}, P_{1} \times P_{4}$ and $P_{3} \times P_{4}$ in the $F_{2}$. Inbreeding depression values were positive in the crosses $P_{1} \times P_{2}, P_{2} \times P_{1}, P_{3} \times P_{1}, P_{4} \times P_{1}, P_{2} \times P_{3}$ and $P_{4} \times P_{3}$ indicating the importance of dominance genes controlling the inheritaance of the character. These results agree with the findings of Deore et al. (1997) on and Zhao et al. (1997). On the other hand, the inbreeding depression values were negative in $P_{1} \times P_{3}, P_{1} \times P_{4}, P_{3} \times P_{2}, P_{2} \times P_{4}, P_{4} \times P_{2}$ and $P_{3} \times P_{4}$ indicating the control of additive gene effect in the inheritance of leaf area for these crosses (Table 2). Assumptions of no epistasis, no multiple allele and uncorrelated distribution were found valid by calculating regression coefficient (b) of covariance between all offspring of the parent and their non - recurrent parent on the variance of these offspring, b is not significantly different than 1.0 in both $F_{1}$ and $F 2$ ( $b=0.40 \pm 0.28,0.84 \pm 0.42$ respectively) indicating the validity of the three assumptions mentioned earlier. This result was also confirmed with Wr , Vr graph as shown in Fig. 4, where Wr was related to Vr by straight line. With regard to the other assumptions of diploid segregation, homozygous parents, they were found valid and no reciprocal differences assumption may be considered valid with some degrees of confidence. In the $F_{1}$, the "E" component estimating environmental effects was significant as shown in Table 3 indicating the important role of environment in the control of leaf area, which agreed with the conclusion of Faluyi (1986) on cashew. The " D " and " $\mathrm{H}_{1}$ " components were not significant. The large environmental role might be expected to mask any heritable variation (Randall and Ruth, 1993). Other ratios couldn't be calculated because the relevant components were not significant (Hayman, 1954). In the $F_{2}$, the " $D$ " component, estimating additive gene effect, was not significant, while " $\mathrm{H}_{1}$ " component, estimating dominance gene effect, was significant indicating that the dominance gene effect played a major role in the inheritance of leaf area. This agreed with Yadav et al. (1981) on wheat. There is a symmetry of positive and negative genes in the parents from the $\mathrm{H}_{2} / 4 \mathrm{H}_{1}$ ratio supported by $\mathrm{KD} / \mathrm{KR}$ ratio, which was larger than unity indicating that the parents seemed to carry more dominant genes than recessives. The proportion $\left(\mathrm{H}_{1} / \mathrm{D}\right)^{1 / 2}$ was larger than the unity indicating the presence of overdominance. This was confirmed with the $\mathrm{Wr}, \mathrm{Vr}$ graph (Fig. 4), where the regression line intercepted Wr in negative position and the degree of dominance could be estimated from the graph in the $F_{1}$, where the regression line intercepted Wr in negative position indicating the presence of overdominance. It can be concluded from the graph that $P_{1}$ had the most recessive genes in $F_{1}$ and $F_{2}$, but $P_{4}$ had the most dominant genes followed by $P_{2}$ and $P_{3}$ in the $F_{1}$, while $P_{4}$ seemed to carry
J. Agric. Sci. Mansoura Univ., 25 (10), October, 2000.

Fig3+4
more recessive genes than dominant. The correlation coefficient between parental mean Yr and Wr , Vr had a negative value in the $\mathrm{F}_{1}$ and $\mathrm{F}_{2}$ indicating that most of dominant genes had positive effect and increased leaf area.

Heritability in broad sense was moderate in $F_{1}(0.52)$ reflecting the environmental effect, while it was high in the $F_{2}(0.89)$ indicating that leaf area is a genetically controlled character (Table 3). Heritability in narrow sense was low ( 0.1 in the $F_{2}$ ) reflecting the week effect of additive genes. The ratio $h_{2} / \mathrm{H}^{2}$ was lower than the unity indicating that leaf area is under the control of one group of genes exhibiting dominance.

## REFERENCES

Abdel-Sabour, M.S.; A.M. Hassan; A.A. Abdel-Shafi; A.S. Sherif and A.A. Hamada (1996). Genetic analysis of diallel cross in bread wheat under different environment conditions in Egypt. Indian Journal of Genetics and plant Breeding 56, (1): 49-61.
Ahmed, M.A. and T.A. Ismail (1999). Gene action in Pea and esterase isozyme activity and its association to heterosis. Zagazig J. Agric. Res., 26 (5): 1281-1292.
Bakheit, B.R. and E.M. Ezzat (1987).Gene action in the inheritance of earliness and some morphological traits in sesame. Assiut Journal of Agricultural Sciences, 81 (1): 175-188.
Cinsoy, A.S. (1992). Inheritance of various quantitative traits in soybean. Soya Fasulye sinde bazi Kantitatif Karaketerlerin Kalitiml Anadolu, 2 (2): 1430. ( C.F. Plant Breeding Abst., 1993. 63 : 10728).

Deore, G.N.; G.N.; B.B. Pawar and C.B. Salunke (1997). Heterosis for Physiological traits and grain yield in pearl millet (Pennisetum americanum L.). Annals of Plant Physiology, 11 (1) : 20-25.
El-Gamasy, A.M. and M.K. Nada (1974). Effect of planting dates on the growth, flowering and seed production of some winter flowering annuals. Agric. Res. Rev., 52(3). 207-225.
El-Torky, M. (1981). A breeding study on Antirrhinum Spp. M.Sc. Thesis, Faculty of Agriculture. Alexandria University.
Faluyi, M.A. (1986). Investigation on seedling vigour in cashew (Anacardium occidentale L.). Plant Breeding, 97: 237-245.
Hayman, B.I. (1954). The theory and analysis of diallel crosses. Genetics, 39: 789-809.
$\ldots \ldots . . . . . . . .$. (1957). Interaction, heterosis and diallel crosses. Genetics, 42: 336-355.
Jinks, J.L. (1954). The analysis of continuous variation in a diallel cross of Nicotiana rustica varieties. Genetics, 39: 797-788.
Khar, A.; N.K. Pathania and S. Navinder (1997). Variability and heritability studies on late cauliflower (Brassica oleracea var. Botrytis L.). Annals of Biology (Ludhiana), 13(1): 127-130.
Kumar, A.A.; M. Ganesh and P. Janila (1998). Combining ability analysis for yield and yield contributing characters in sunflower (Helianthus annus L.). Annals of Agricultural Research,19(4):437-440.

Madić, M. (1996). Inheritance of spike traits and grain yield in barley (Hordeum vulgare L.). hybrids. Review of Research Work at the Faculty of Agriculture, Belgrade, 41. (1): 53-65. (C.F. Plant Breeding Abst., 1997. 67 : 8030).

Mahdy, E.E.; H.H. El-Hinnawy and M.M. Abdalla (1983). Gene action in the inheritance of earliness and plant height of cotton. Assiut Journal of Agricultural Sciences, 14(2): 273-285.
Mather, K. and J. L. Jinks (1971). Biometrical genetics. Chapman and Hall, LTD. London, $382-416$.
Misiha, A. (1991). Inheritance of some quantitative characters in Antirrhinum majus L. Bull. Fac. of Agric. Univ. of Cairo, 42 (1): 217-236.
Randall, J. M. and G. S. Ruth (1993). Heritability of floral traits for the perennial wild flower Penstemon centranthifolius (Scrophulariaceae) clones and crosses. Heredity, 71: 185-192.
Rastogi, K. B.; B. N. Korla; A. K. Joshi and M.C. Thakur (1995). Variability studies in chinese cabbage (Brassica chinensis, L.). Advances in Horticulture and Forestry, 4: 101-107.
Shamsuddin, A. K. M.; M. A. Newaz and C. A. Razzague (1980). Genetic analysis of leaf yield and component characters in Tabacco (Nicotiana tabacum L.). Plant Breeding, 84: 139-147.
Singh, P.K. and B.D. Chaudri (1977). Biometrical methods in quantitative genetic analysis. Kalyani Publlishers, Ludhiana, Newdelhy.
Singh, S.P. and S. Sudhir (1996). Genetics of developmental traits in opium poppy (Papver somniferum L.). Recent Horticulture, 3 (1): 122-125.
Steel, R.G. D. and Torrie, J. H. (1986). Principles and procedures of statistics: A biometrical approach. Second Edition. Mc. Graw-Hill. Book Company.
Weber, W. E. (1976). A ${ }_{10}$-Parent diallel for quantitative genetic studies in peas (Pisum sativum L.). Plant Breeding, 77: 30-42.
Yadav, V.S. and B.S. Chankar (1991). Genetic components for yield and its contributing characters in okra. Indian Journal of Horticulture, 48(1): 7174.

Yadav, B.; Y. Mohd and L. Rai. (1981). Genetic studies on some traits above the flag leaf node in wheat. Plant Breeding, 86: 305-310.
Zhao, M.; M.Y. Wang and S. Li (1997). Comparison of main photosynthetic characteristics in hybrids and parents of maize (Zea maize L.). Acta Agriculturae Boreali Sinica, 12 (2): 39-43.

# تحليلات بيومترية لبعض الصفات الكمية الهامة في حنك اللسبع <br> ا. مواصفات النمو الخضري. <br> مصطفى بدر* ، محمد جمال التركي* ، رابحة عباس** وجيهان جابر*.. <br> * قسم الزهور ونباتات الزينة كلية الزراعة، جامعة الإسكندرية. <br> ** معهُ بحوث البساتين، مركز البحوث الزراعة، الإسكندرية. 


 التنحكم فى وراثة صفات النمو الخضري وكذلك الفعل الجينى المتحكم فى الصفات الكمية المدروسة لتحديد إمكانية إنتاج الصفات الخضرية المر غوبة تبعا لنتطبات السوق.
P1 = Sonnet wit وقد استخمت فى الار اسة أربعه أصناف من نبات حنك السبع وهم (white),
$\mathrm{P}_{2}=$ Sonnet karmijn (red), $\mathrm{P}_{3}=$ Sonnet rose (rose) and $\mathrm{P}_{4}=$ Sonnet geel (yellow)
حيث أجريت بينهم كل التهجينات المحكنة. وأظهرت النتائج وجود فروق معنوية بين الهجن فى كل الصفات المدروسة وقت استخدمت طريقة هايمان لتحديد النظام الوراثي التّحكم في ور اثة الصفات.

بالنسبة لصفة طول النبـت : ثبتت أهية الفعل الجينى الهضيف والسيادى فى كلا الجيلين مع وجود سيادة متنوقة. درجة التوريث بالمعنى الضيق متوسطة ولم يظهر إنخفاض لطول النبات فى الجيل الثاني عن الجيل الأول لأغلب الهجن ولذلك يككن تحسين الصفة بالإنتخاب. اظظهر التحليل اليباني أن الأب ( على معظم الجينات السائّدة كما أن معظم الجينات السائدة تؤدى إلى زيادة طول النبات ـ ظهرت قوة الهجين فىى كل الهجن الناتجة.

وبالنسبة لصفة عدد الأفرع : ثبتّت أهية التأثير المضيف للجينات. درجة التوريث بالمنى الضيق كانت متوسطة ممـا يغى إمكانية تحسين الصفة بالانتخاب. معظم الجينـات السانـأدة تقلل عدد الأفرع والأب (P1) يحتوى على معظم الجينات المتنحية. وجود سيادة متنوقة فى الجيل الأول وسيادة جزئية فى الجيل الثاني. الجينات السائدة اكثر تكرارا من الجينات المتنحية وظهرت قوة الهجين فى اغلب الهجن.

أما بالنسبة لصفة عدد الأوراق : فلقت ثّتت أهية كل من الثأثير المضيف للجينات والتأثيّير السيادى مع وجود سيادة متنوقة فى كال الجيلين ووجود التنوق فى الجيل الثاني. درجة التُوربث بالمعنى الضيق متوسطة. الجينـات


الدتنحية هي التي تقلل الصفة فى الجيل الثاني. الأبوين (P) و و (P3 ) يحتويان على أغلب الجينات السائثة.
و عثد دراسة المساحة الورقية : كانت النأثيرات البيئية معنوية بينما التأثير ات الور اثية غير معنوية فى الجيل الأول. اتضحت أهمية الفعل الجينى المضيف فى الجيل الثاني وكذلك وجود السيادة المتفوقة. احتوت الآباء
 الأب (P2 ) على معظم الجينات السائّة.

Table1: Mean values of plant height, number of branches/ plant, number of leaves/ plant and leaf area of the $F_{1}$ and $F_{2}$ generations for the different selfings and crosses of Antirrhinum majus.

| Genotypes ${ }^{1)}$ | Mean ${ }^{2}$ plant height (cm) |  |  |  | Mean ${ }^{2}$ no. branches/plant |  |  |  | Mean ${ }^{2}$ no. leaves/ plant |  |  |  | Mean ${ }^{1 /}$ leaf area ( $\mathrm{cm}^{2}$ ) |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | $\mathrm{F}_{1}$ |  | $F_{2}$ |  |  |  |  |  | $\mathrm{F}_{1}$ |  | $F_{2}$ |  | $\mathrm{F}_{1}$ |  | $\mathrm{F}_{2}$ |  |
| $\mathrm{P}_{1} \times \mathrm{P}_{1}$ | 35.4 | ghi | 48.9 | gh | 82.3 | a | 66.3 | cd | 1200.6 | a | 1123.3 | bc | 7.2 | d | 7.90 | i |
| $\mathrm{P}_{2} \times \mathrm{P}_{2}$ | 33.7 | hi | 51.5 | fgh | 56.3 | bc | 71.3 | bc | 770.6 | h | 1003.3 | cd | 8.8 | cd | 10.7 | ghi |
| $\mathrm{P}_{3} \times \mathrm{P}_{3}$ | 45.5 | bcde | 61.9 | cd | 33.3 | d | 20.3 | h | 701.0 | fgh | 766.0 | fg | 9.8 | bcd | 13.5 | defg |
| $\mathrm{P}_{4} \times \mathrm{P}_{4}$ | 32.7 | i | 41.1 | i | 31.0 | d | 26.6 | gh | 606.6 | gh | 652.6 | g | 11.6 | bcd | 9.30 | hi |
| $\mathrm{P}_{1} \times \mathrm{P}_{2}$ | 38.8 | fgh | 53.0 | fg | 81.6 | a | 90.2 | a | 1084.0 | ab | 1377.3 | a | 12.3 | bcd | 12.0 | efgh |
| $\mathrm{P}_{2} \times \mathrm{P}_{1}$ | 40.7 | defg | 58.7 | d | 88.3 | a | 83.9 | ab | 985.0 | abcd | 1203.6 | b | 13.7 | bc | 14.1 | cde |
| $\mathrm{P}_{1} \times \mathrm{P}_{3}$ | 49.3 | ab | 64.6 | bc | 56.0 | bc | 36.3 | efgh | 896.3 | bcdef | 991.0 | cde | 11.1 | bcd | 16.2 | bcd |
| $\mathrm{P}_{3} \times \mathrm{P}_{1}$ | 51.6 | a | 69.0 | a | 49.0 | cd | 37.8 | efg | 575.0 | h | 839.0 | ef | 20.6 | a | 20.0 | a |
| $\mathrm{P}_{1} \times \mathrm{P}_{4}$ | 41.1 | g | 58.5 | de | 32.6 | d | 45.0 | ef | 587.0 | h | 913.3 | def | 12.9 | bc | 16.8 | bc |
| $\mathrm{P}_{4} \times \mathrm{P}_{1}$ | 44.5 | f | 62.8 | bcd | 48.3 | cd | 49.8 | de | 592.6 | h | 1112.0 | bc | 13.2 | bc | 13.7 | def |
| $\mathrm{P}_{2} \times \mathrm{P}_{3}$ | 45.3 | bcde | 61.1 | cd | 47.3 | cd | 47.3 | ef | 1030.3 | abc | 1005.6 | cd | 12.4 | bcd | 12.7 | efg |
| $\mathrm{P}_{3} \times \mathrm{P}_{2}$ | 46.8 | abc | 48.3 | h | 74.0 | ab | 50.5 | de | 974.6 | bcde | 1131.0 | bc | 9.5 | bcd | 10.9 | fgh |
| $\mathrm{P}_{2} \times \mathrm{P}_{4}$ | 45.5 | bcde | 59.1 | d | 57.3 | bc | 48.5 | e | 806.3 | defg | 1018.0 | cd | 14.5 | b | 16.0 | bcd |
| $\mathrm{P}_{4} \times \mathrm{P}_{2}$ | 39.8 | efg | 54.3 | ef | 62.0 | bc | 46.3 | ef | 766.0 | efgh | 1135.0 | bc | 10.6 | bcd | 13.6 | defg |
| $\mathrm{P}_{3} \times \mathrm{P}_{4}^{2}$ | 48.8 | ab | 63.8 | bc | 56.6 | bc | 36.8 | efgh | 858.3 | cdef | 774.0 | fg | 11.5 | bcd | 17.8 | ab |
| $\mathrm{P}_{4} \times \mathrm{P}_{3}$ | 46.2 | abcd | 67.1 | ab | 46.6 | c | 31.5 | fgh | 600.6 | h | 877.0 | def | 13.3 | bc | 13.6 | defg |
| L.S.D. 0.05 |  |  |  |  |  |  |  |  | 21 |  |  |  |  |  |  |  |

1) Seed parent is the first one, $P_{1}=$ white, $P_{2}-$ red- purple, $P_{3}=$ red, $P_{4}=$ yellow. N.S; *** Not significant, significant at 0.05 and 0.01 respectively.
2) Values in the same column not followed by the same letter are significantly different at the $5 \%$ probability level.

Table 2 : Estimates of heterosis and inbreeding depression (I.D) for plant height, number of branches/ plant, number of leaves/ plant and leaf area for the different crosses of Antirrhinum majus.

| Genotypes ${ }^{1{ }^{1}}$ | plant height (cm.) |  | No. of branches/ plant |  | No. of leaves/ plant |  | leaf area (cm²) |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Heterosis | I.D. | Heterosis | I.D. | Heterosis | I.D. | Heterosis | I.D. |
| $\mathrm{P}_{1} \times \mathrm{P}_{2}$ | $4.20{ }^{\text {N.S }}$ | $-0.50{ }^{\text {N.S }}$ | $+12.3{ }^{\text {N.S }}$ | $-16.1^{\text {N.S }}$ | +98.4 ${ }^{\text {N.S }}$ | -20.9 ${ }^{\text {N.S }}$ | 4.3 d.S | +0.5** |
| $\mathrm{P}_{2} \times \mathrm{P}_{1}^{2}$ | 6.10 * | -9.80 ** | +19.0* | $-0.22^{\text {N.S }}$ | $-0.6{ }^{\text {N.S }}$ | -15.4 ${ }^{\text {N.S }}$ | $5.7 *$ | +4.50* |
| $\mathrm{P}_{1} \times \mathrm{P}_{3}$ | 8.80** | -2.40 N.S | -1.8 N.S | +26.9** | $-54.50^{\text {N.S }}$ | $-3.1{ }^{\text {N.S }}$ | $2.6{ }^{\text {N.S }}$ | -36.9** |
| $\mathrm{P}_{3} \times \mathrm{P}_{1}$ | 11.10** | $-6.50{ }^{*}$ | $-8.8{ }^{\text {N.S }}$ | $+13.4^{\text {N.S }}$ | $-375.8^{* *}$ | $-34.3{ }^{\text {N.S }}$ | 12.1** | +7.70** |
| $\mathrm{P}_{1} \times \mathrm{P}_{4}$ | 7.00** | -8.20** | -24.05** | -52.1** | -316.6** | $-44.2{ }^{\text {N.S }}$ | $3.5{ }^{\text {N.S }}$ | -22.4** |
| $\mathrm{P}_{4} \times \mathrm{P}_{1}$ | $10.4{ }^{* *}$ | -9.60 ** | $-8.3^{\text {N.S }}$ | $-12.6{ }^{\text {N.S }}$ | -311.0** | $-76.4{ }^{\text {N.S }}$ | 3.8 N.S | $+3.70{ }^{\text {N.S }}$ |
| $\mathrm{P}_{2} \times \mathrm{P}_{3}$ | $5.70{ }^{*}$ | $-3.90^{\text {N.S }}$ | $+2.5{ }^{\text {N.S }}$ | $-9.7{ }^{\text {N.S }}$ | +294.8** | +8.8 ${ }^{\text {N.S }}$ | $3.1{ }^{\text {N.S }}$ | +5.60** |
| $\mathrm{P}_{3} \times \mathrm{P}_{2}$ | $7.20{ }^{* *}$ | +26.7** | +29.2** | +25.5** | $+238.8{ }^{*}$ | $-9.2{ }^{\text {N.S }}$ | $0.2{ }^{\text {N.S }}$ | $-4.20 *$ |
| $\mathrm{P}_{2} \times \mathrm{P}_{4}$ | 12.30** | $+0.80 \mathrm{N.S}$ | +13.6 ${ }^{\text {N.S }}$ | $+7.3^{\text {N. }}$ | +117.7 N.S | $-17.9^{\text {N.S }}$ | $4.3{ }^{\text {N.S }}$ | -3.40 N.S |
| $\mathrm{P}_{4} \times \mathrm{P}_{2}$ | $6.60{ }^{*}$ | $-1.20{ }^{\text {N.S }}$ | +18.3** | +17.9** | +77.4 ${ }^{\text {N.S }}$ | $-13.3{ }^{\text {N.S }}$ | $0.4{ }^{\text {N.S }}$ | $-18.80{ }^{* *}$ |
| $\mathrm{P}_{3} \times \mathrm{P}_{4}$ | 9.70** | $-2.00{ }^{\text {N.S }}$ | +24.4** | +26.8** | +204.5* | +17.5 ${ }^{\text {N.S }}$ | $0.8{ }^{\text {N.S }}$ | -46.0** |
| $\mathrm{P}_{4} \times \mathrm{P}_{3}$ | 7.10** | -14.90** | +14.4 ${ }^{\text {N.S }}$ | +22.5* | $-53.2{ }^{\text {N.S }}$ | -34.9 ${ }^{\text {N.S }}$ | $2.6{ }^{\text {N.S }}$ | +5.2* |
| L.S.D. 0.05 | 5.2 | 5.2 | 16.4 | 17.9 | 187.3 | 191.9 | 4.6 | 4.2 |
| L.S.D. 0.01 | 7.0 | 7.1 | 22.1 | 24.2 | 252.2 | 258.5 | 6.2 | 5.7 |

1) Seed parent is the first one, $P_{1}=$ white, $P_{2}$ - red- purple, $P_{3}=$ red, $P_{4}=$ yellow.
N.S; ${ }^{* * *}$ Not significant, significant at 0.05 and 0.01 respectively.
2) values in the same column not followed by the same letter are significantly different at the $5 \%$ probability level.

Table 3 : Estimates of genetic parameters and ratios with their respective standard errors in $F_{1}$ and $F_{2}$ for plant height, number of branches/ plant, number of leaves/ plant and the leaf area of Antirrhinum majus.

| Estimate | Plant height (cm.) |  | No. of branches/ plant |  | No. of leaves/ plant |  | Leaf area ( $\mathrm{cm}^{2}$.) |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | $\mathrm{F}_{1}$ | $\mathrm{F}_{2}$ | $\mathrm{F}_{1}$ | $\mathrm{F}_{2}$ | $\mathrm{F}_{1}$ | $\mathrm{F}_{2}$ | $\mathrm{F}_{1}$ | $\mathrm{F}_{2}$ |
| D | $30.38 \pm 1.68^{*}$ | $71.85 \pm 10.47^{*}$ | $530.92 \pm 67.06^{*}$ | 659.95 $\pm 27.41^{*}$ | 63394.11さ9207.80* | 43077.74土1552.65* | $00.05 \pm 3.15^{\text {N.S }}$ | $4.61 \pm 2.86^{\text {N.S }}$ |
| $\mathrm{H}_{1}$ | $63.27 \pm 4.89^{*}$ | $728.15 \pm 125.01^{*}$ | $543.37 \pm 194.94 *$ | $499.89 \pm 318.71^{\text {N.S }}$ | 105183.52 $\pm 26766.05^{*}$ | 136704.67 $\pm 18528.87^{*}$ | $18.03 \pm 9.18^{\mathrm{N.S}}$ | $150.28 \pm 33.26^{* *}$ |
| $\mathrm{H}_{2}$ | $64.15 \pm 4.52^{*}$ | $610.12 \pm 112.44^{*}$ | $440.44 \pm 179.94^{*}$ | $615.54 \pm 294.20^{\text {N.S }}$ | $71716.33 \pm 24707.12^{*}$ | 118553.36 $16664.84^{*}$ | $16.37 \pm 8.47^{\text {N.S }}$ | $134.67 \pm 30.70^{*}$ |
| $\mathrm{h}^{2}$ | $140.95 \pm 3.06{ }^{*}$ | $159.80 \pm 76.27^{*}$ | $97.06 \pm 122.05^{\text {N.S }}$ | $-394.19 \pm 199.55^{\text {N.S }}$ | $-415.08 \pm 16758.45^{\text {N.S }}$ | $8560.72 \pm 11303.50^{\text {N.S }}$ | $26.67 \pm 5.74 *$ | $36.50 \pm 20.82^{\text {N.S }}$ |
| F | $-6.28 \pm 4.32^{\text {N.S }}$ | $120.65 \pm 53.08^{*}$ | $231.35 \pm 172.28^{\text {N.S }}$ | $-315.05 \pm 138.89^{\mathrm{N.S}}$ | 57148.04 $\pm 23655.27^{*}$ | $-10887.18 \pm 7867.38^{\text {N.S }}$ | $2.45 \pm 8.11^{\text {N.S }}$ | $9.14 \pm 14.49^{\mathrm{NSS}}$ |
| E | $4.33 \pm 0.75^{*}$ | $2.4 \pm 4.6^{\text {N.S }}$ | $43.1 \pm 29.99^{\text {N.S }}$ | $35.93 \pm 12.25^{\text {N.S }}$ | $5610.13 \pm 4117.85^{\text {N.S }}$ | $3234.83 \pm 694.36^{*}$ | $3.4 \pm 1.41^{*}$ | $1.1 \pm 1.27^{\text {N.S }}$ |
| $\left(\mathrm{H}_{1} / \mathrm{D}\right)^{1 / 2}$ | 1.44 | 1.59 | 1.01 | 0.43 | 1.2 | 0.89 | 1.95 | 2.85 |
| $\mathrm{H}_{2} / 4 \mathrm{H}_{1}$ | 0.25 | 0.20 | 0.20 | 0.30 | 0.17 | 0.21 | 0.24 | 0.22 |
| KD/KR | 0.86 | 3.23 | 1.54 | 1.00 | 2.07 | 0.75 | 2.11 | 2.06 |
| $\mathrm{r}\left(\mathrm{y}_{\mathrm{r}}, \mathrm{W}_{\mathrm{r}}+\mathrm{V}_{\mathrm{r}}\right)$ | -0.93 | -0.69 | 0.09 | 0.92 | 0.99 | -0.51 | -0.91 | -0.81 |
| $\mathrm{h}^{2}$ Ns | 0.46 | 0.35 | 0.56 | 0.60 | 0.45 | 0.45 | 0.12 | 0.10 |
| $\mathrm{h}^{2}$ Bs | 0.88 | 0.95 | 0.87 | 0.86 | 0.87 | 0.86 | 0.52 | 0.89 |
| $\mathrm{h}^{2} / \mathrm{H}_{2}$ | 2.19 | 0.26 | 0.22 | 0.64 | 0.05 | 0.07 | 0.35 | 0.27 |

N.S., * : Non significant and significant respectively (The significance was defined in the $F_{1}$, when the value exceeded 1.96; while in the $F_{2}$, the significance was tested by t- test at $\mathrm{P}=0.05$ and 2 degrees of freedom).

