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## Influence of some Biofertilizers and Chemical Fertilizers on some Vegetative and Root Growth Characteristics of *Casuarina equisetifolia* Seedlings Grown in Sandy Soil

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### ABSTRACT

A pot experiment was carried out at the Experimental Farm and Laboratory of Horticulture Department, Faculty of Agriculture, Benha University Egypt during the 2019 and 2020 successive growing seasons. The study was carried out to define the efficiency of various biofertilizers Minia Azotein (MA), Frankia and Vesicular Arbuscular Mycorrhiza. each alone or combined with two doses of chemical fertilizer NPK at 75% and 50% from the recommended dose (350:200:100 kg/ fed) and/or the full dose of 100% NPK for improving some vegetative and root growth characteristics of *Casuarina equisetifolia* seedlings grown in sandy soil. Results showed that all treated seedlings of casuarina with Frankia 50g/ plant, and Vesicular Arbuscular Mycorrhiza (VAM) 50 ml/plant in combination with 75% NPK T8 and T10 and or NPK 100% recommended dose (10:5:2.5g/plant) T2 recorded significant increases in plant height, stem diameter, number of branches, biomass fresh and dry weights compared to the other treatments and control. The highest rooting values of root length, root fresh, and dry weights of casuarina plants were found with the abovementioned treatments followed by Minia Azotein (MA) combined with 75%NPK (T6) in both seasons while the other treatments did not attain any significant variances in vegetative and root growth parameters. It can be concluded that Frankia 50g/plant, Vesicular Arbuscular Mycorrhiza (VAM) 50 ml/plant with 75% of the recommended dose (7.5:3.75:1.875g/plant)T6, T8, and/ or 100% of the recommended dose of NPK (10:5:2.5g/plant) T2 were the best in improving vegetative and root characteristics of *Casuarina equisetifolia* seedlings.

**Keywords:** *Casuarina equisetifolia*, Minia Azotein (M.A), Frankia, Arbuscular Mycorrhiza Fungi (AMF), chemical fertilizer NPK, vegetative and root growth.

### INTRODUCTION

*Casuarina* is a needle tree an upright, fast-growing, evergreen tree with many branches. native to Asia where they call it (Jemara or Simara) and a length of up to 35m they multiplied by seeds belonging to the family (Casuarinaceae) coexisting with radial root nodules (Karthikeya, 2016). These trees tolerate drought, salinity, and heavy metal pollution (Pooja and Katharina, 2021) including three species, and their hybrids of the three species, *C. equisetifolia*, and *C. glauca* are highly salt-tolerant fall with strong winds (Wheeler *et al.*, 2011). *Casuarina equisetifolia* is an important tree and suffers from bacterial wilt caused by *Ralstonia solanacearum* (Yongcheng *et al.*, 2021). It acts as a shelter for ants, bees, and birds (Alok, 2021). *Casuarina equisetifolia* has traditionally been used to treat neurological problems, acne, sore throats, stomach ulcers, constipation, cough, diabetes, and dysentery (Vain *et al.*, 2022). It helped to improve the ecosystems in agricultural systems and helps reduce negative environmental impacts such as erosion, runoff, evaporation, and increases soil organic matter (Ian, 2021). It has the ability to treat wastewater by removing the methylene blue dye from it (Helly *et al.*, 2021). The actinorhizal plant *Casuarina equisetifolia* L. has a mutualistic relationship with Frankia and promotes nitrogen fixation in root nodules (Yujin *et al.*, 2021).

The role of N<sub>2</sub>-fixing bacteria has been demonstrated by several researchers (Dadarwall *et al.*, 1997; Hedge *et al.*,

1999; Hauwaka, 2000). N<sub>2</sub>-fixing microorganisms have a function of fixing N<sub>2</sub> in the atmosphere, their effect on photosynthetic hormones such as indole acetic acid, gibberellins, and cytokinin, and their role in promoting the absorption of many nutrients while improving water quality, which enhances the characteristics of vegetative growth. Anti-bacterial and anti-fungal protect the host plants from infection (Soliman, 2019).

For N fixation, Frankia relates to *Casuarina equisetifolia*, and it is predicted that Frankia fixes up to 362 kg N/ha./Year, which is an important nutrient for all plant metabolic processes and development (Shantharam and Mattoo, 1997 and Ojha *et al.*, 2018). N<sub>2</sub> fixation can be catalyzed by Frankia by adding nitrogen at the recommended rate, actinorhizal plants may fix nitrogen in a symbiotic manner (Mackay *et al.*, 1987). In addition to Mycorrhiza and the nitrogen-fixing bacteria Frankia, actinorhizal plants are also able to form a unique type of roots called "proteoid roots" or "cluster roots" in response to the detrimental effects of nutrient deficiencies in the soil (Malajczuck and Lamont, 1981; Crocker and Schwintzer, 1993).

Vesicular Arbuscular Mycorrhiza (VAM)) secrete the enzyme phosphatase that dissolves organic phosphorous and works to increase the activity of phosphorous-dissolving bacteria, which helps to increase the amount of it in the soil (Yang *et al.*, 2022). The fungus absorbs phosphorous through a variety of mechanisms, including the secretion of the

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enzyme Phosphates by the fungus hyphae, which works to dissolve organic phosphorous and convert it into forms suitable for plant absorption, and the secretion of hydroxyl acids, which seize the elements calcium, iron, and aluminum while leaving the phosphorous element dissolved in the soil solution. Because of their mutually beneficial relationship, mycorrhizal fungi increase the activity of phosphate-dissolving bacteria. As a result, the amount of dissolved phosphorus in the soil solution rises (Dinkelaker and his associates, 1995).

Mineral NPK fertilizer promotes the vegetative growth, root development, flower development, seed development, and fruit development of tree seedlings. Because nitrogen is found in amino acids, proteins, alkaloids, coenzymes, and certain vitamins, it plays an important role in metabolism, heredity, reproduction, and growth, according to experts (James and Michael, 2009). Trees need nutrients in order to carry out the various biological processes inside them. Nitrogen enters the formation of proteins, amino acids, enzymes, nucleic acids, RNA, DNA, chlorophyll, vitamins, and alkaloids (Curtis, 1996). Nitrogen is absorbed in the form of nitrate ions NO<sub>3</sub> or ammonia NH<sub>4</sub> (Anthony and Glass, 2010). A moving element that shows symptoms of deficiency on the lower leaves, the plant needs it by 1-5%. Its absence hinders the growth of the plant by exceeding the permissible limit and the root total is more than the vegetative (Arnold, 2015). Moreover, phosphorous enters the formation of phospholipids and nucleic acids, in addition to its role in the processes of oxidation, reduction, photosynthesis and respiration (James, 2020). In addition to its entry into the synthesis of ATP, the carrier of energy, it is absorbed in the form of mono- and di-phosphate, which is a mobile element. Symptoms of deficiency appear on the aged leaves (Wei *et al.*, 2021). Potassium is found in the plant in the form of soluble inorganic salts. It has a role as an activator of enzymes, the representation of carbohydrates and proteins, and the opening and closing of stomata (Kaan and Ramazan, 2022). Chemical fertilizers have much damage to humans, plants, soil, and the environment, including what is caused by an increase in nitrogen fertilizers, from the conversion of nitrates to nitrites that cause bowel cancer and the increase in the incidence of pests, insects, and diseases in plants and the accumulation of heavy elements in the soil beside the polluting soils of the environment resulting from the phosphate industry. (Nikita and Puneet, 2020 and Masindi and Muedi, 2018).

The research aims to define the efficiency of various biofertilizers Minia Azotein (MA), Frankia and vesicular Arbuscular Mycorrhiza (VAM) each alone or combined with tow doses of chemical fertilizer NPK at 75% and 50% from the recommended dose (350:200:100 kg/fed) and / or the full dose of 100% NPK for improving some vegetative characteristics i.e. plant height, stem diameter, number of branches, biomass fresh and dry weights , and root growth characteristics i.e. root length, root fresh and dry weights of *Casuarina equisetifolia* seedlings grown in sandy soil.

## MATERIALS AND METHODS

This study was carried out at the Experimental Farm and Laboratory of Horticulture Department, Faculty of Agriculture, Moshtohor, Benha University, Egypt, during two successive seasons of 2019 and 2020 on *Casuarina*

*equisetifolia* seedlings to evaluate the effectiveness of biofertilizer Minia Azotein(MA), Frankia and Vesicular Arbuscular Mycorrhizal (VAM)) each alone or combined with two doses of chemical fertilizer NPK at 75% and 50% of the recommended dose (350:200:100 kg/fed ) and or the full dose of 100% NPK for improving some vegetative growth and root growth characteristics i.e. plant height, stem diameter, number of branches, biomass fresh and dry weights, root length, root fresh and dry weights of *Casuarina equisetifolia* seedlings grown in sandy soil. seedlings of one-year-old with an average height of 10-15 cm diameter of 3-6mm were purchased from EL- Orman Garden, Ministry of Agriculture, Egypt. On March 1<sup>st</sup> week for both experimental seasons, the seedlings were repotted in 40cm plastic pots filled with 30kg of sandy soil, one seedling in each pot.

Physical and chemical properties of the used soil were done according to the method described by Okalebo *et al.* (2002) and listed in Table (a)

**Table a. Physical and chemical analysis of the experimental soil before the application of any fertilizers in both seasons 2019 and 2020.**

Soil analysis	2019	2020
Sand %	81.04	80.92
Silt %	12.28	11.83
Clay %	6.86	25.7
Soil texture	Sandy	Sandy
Organic matter %	0.11	0.09
CaCO <sub>3</sub>	3.35	3.51
pH (1:2.5)	8.01	7.92
E. C (m mhos /cm)	1.23	1.19
Total N (%)	0.001	0.001
Available P (mg/kg)	3.00	2.50
Available K (mg/kg)	94.00	105.00
Fe (mg/kg)	10.10	9.80
Mn (mg/kg)	8.50	8.30
Cu (mg/kg)	1.10	0.90
Zn (mg/kg)	0.88	1.00

**Biofertilizer:- 1- Minia Azotein MA (commercial name)** was used. It contains live cells of N-fixing bacteria (1 ml = 10<sup>7</sup> cells of bacteria), were obtained from the Laboratory of Biofertilizers, Department of Genetic, Fac. of Agric., Minia University according to Abdou *et al.* (2006) The three biofertilizers were used at the concentrations of 50 ml/pot of Minia Azotein MA, 50g/ pot of Frankia, and 50ml/pot of Vesicular Arbuscular Mycorrhiza VAM, three times on 16<sup>th</sup> April, 16<sup>th</sup> June, and 16<sup>th</sup> August one week after chemical fertilizer treatments had been done and immediately after irrigated pots with 75% of its field capacity( 2.25L/pot) in both seasons of 2019 and 2020.

**2- Frankia the double layer technique** was used to separate Frankia sp. from the collected *Casuarina* root nodules (Murry *et al.*, 1984). The chosen nodules were washed several times in water to remove most dirt and organic material adhered to the surface, then thoroughly cleaned under the dissecting microscope to remove any soil and organic particles left adhered to the nodule surface or inserted between the lobes. Surface lobes were sterilized in sodium hypochlorite (2.0 percent) and HgCl (0.1 percent in 0.5 percent HC) for 15 minutes, then washed multiple times with sterile distilled water. Individual lobes were grown in nutrient broth for one week at 29+2°C to assess sterilization quality (Carrasco *et al.*, 1992). Using a sterile scalable, the sterilized lobes were sliced into little pieces. The little lobe pieces were eventually distributed over a 1.59 percent agar modified-Q-mod bottom layer in a Petri plate. 3 ml of semi-solid modified Q-medium

was placed on top of the overlayer, covering the lobe pieces (Caru, 1993), and cultured for 2-3 months at 29+2°C. On modified -Q-mod, Frankia colonies that grew out of nodule fragments and had normal hyphae and sporangia were separated (Lalonde and Calvert, 1979). Purification tests were performed on Frankia isolates (Diem and Dommergues 1983). The pure Frankia isolates were kept at 29+2°C on a modified BAP liquid medium, and subculturing was done on the same medium every week.

**3-Vesicular Arbuscular Mycorrhizal (VAM)** was collected from the Biofertilizer Laboratory of Ain Shams University, Genetics Department, (AMF) pollen was originally extracted from the soil around wheat roots in the experimental field of Fac. Agriculture, Ain Shams University, Shubra El-Kheima, Cairo, Egypt, according to Gerdmann and Nicolsan (1963).

Chemical fertilizer N,P and K as recommended dose 350 kg/fed of amonium sulfate (20.6% N), 100 kg/fed of potassium sulfate (48% K<sub>2</sub>O) and 200 kg/fed of calcium superphosphate (15.5%P<sub>2</sub>O<sub>5</sub>) all obtained from Kema factory of Aswan. The NPK doses were applied as dressing inside each pot in three equal portions added at 9<sup>th</sup> April, 9<sup>th</sup> June and 9<sup>th</sup> August at one week interval prior to each biofertilizer treatments. Each pot was received (10:5:2.5g) as recommended dose of NPK and (7.5:3.75:1.875g) as 75% of recommended dose of NPK and (5:2.5:1.25g) as 50% of recommended dose of NPK.

The experiment was arranged in a randomized complete block design (RCBD) with three replicates for each treatment each replicate contained 3 Casuarina seedlings (11 treatments × 3 replicates × 3 seedlings) =99 seedlings every season.

- 1- Control-: without any fertilizer T1.
- 2- 100 % NPK as recommended dose:(10:5:2.5g/ pot) T2.
- 3- Minia Azotein MA: (50ml/pot) T3.
- 4- Frankia: (50g/pot) T4.
- 5- Vesicular Arbuscular Mycorrhizal VAM: (50ml/pot) T5.
- 6- 75% NPK + MA:(50ml/pot) T6.
- 7- 50% NPK + MA:(50ml/pot) T7.
- 8- 75% NPK + Frankia: (50g/pot) T8.
- 9- 50% NPK + Frankia :(50g/pot) T9.
- 10- 75% NPK +VAM: (50ml/pot) T10.
- 11- 50% NPK +VAM: (50ml/pot) T11.

All horticultural practices including irrigation and weed management were done as recommended to this respect.

At the end of each season on November 1<sup>st</sup> in both seasons, the data were recorded on vegetative growth characteristics of casuarina plants i.e. plant height(cm), stem diameter(cm), number of branches, biomass fresh and dry weights(g/plant). On root growth characteristics i.e. root length(cm), root fresh and dry weights (g/plant).

#### **Statistical analysis**

Data were obtained from treatments of the factors under study for analyzes of *variance* (ANOVA) was used to assess the significance of the data at  $P \leq 0.05$  and differences were evaluated using least significant differences (L.S.D) according to (Snedecor and Cochran, 1989) using MSTAT-C statistical software package(1986).

## **RESULTS AND DISCUSSION**

### **Vegetative characteristics**

#### **Plant height (cm):**

The data in Table (1) showed a significant increase in plant height compared to control. The tallest seedlings was

used recorded when mineral NPK fertilizer as a recommended dose (10:7.5:2.5g/pot) was used and attained the highly percentages of increases reached to 47.78 and 51.27 % over control in the first and second season respectively. Followed by treatment 75% of NPK with Frankia (50g/plant) which gave a significant increase in plant height reached to 46.46 and 51.16 % over control in the first and second season respectively, then treatment by Mycorrhizae (50ml/pot) plus 75% NPK, which gave a significant increase in length reached to 46.46 and 51.16 % over control in the first and second season. respectively. According to this data the differences between the highest three treatments had no significant.

#### **Stem Diameter (cm):**

The data in Table (1) showed that the recommended dose of 100% NPK T2 and Frankia (50g/pot) combined with 75% of NPK T8 followed by Mycorrhizae (50ml/pot) combined with 75% of NPK T10 produced the thickest diameter seedlings of Casuarina with non-significant differences between them whereas Minia Azotein (50ml/pot) combined with 75% ranked the fourth in this concern while the other treatments did not attain any significant variances in stem diameter compared to control (00.0) T1.

#### **Number of branches:**

The data in Table (1) showed that the recommended dose (10:7.5:2.5g/pot) of 100% mineral NPK fertilizer T2 and Frankia (50g/pot) combined with 75% of mineral NPK fertilizer (7.5:3.75:1.875g) T8 followed by Mycorrhiza (50ml/pot) combined with 75% of mineral NPK fertilizer T10 produced the highest values of branch number of seedlings of casuarina with non significant differences between them whereas Minia Azotein (50ml/pot) combined with 75% of mineral NPK fertilizer T6 ranked the fourth in this concern while the other treatments did not attain any significant variens in number of branches compared to control (00.0)T1.

#### **Biomass fresh weight(g):**

The data in Table (1) showed a significant increase in biomass fresh weight compared to control. the heaviest weight of biomass fresh weight was recorded when the mineral NPK fertilizer as a recommended dose was used and attained the highly percentages of increases reached to 48.13 and 48.99 % over control in the first and second season respectively. Followed by treatment 75% of NPK with Frankia (50g/plant) which gave a significant increase in biomass fresh weight reached to 45.36 and 46.17 % over control in the first and second season respectively, then treatment by Mycorrhiza (50ml/pot) plus 75% NPK, which gave a significant increase in biomass fresh weight reached to 43.28 and 44.07 % over control in the first and second season respectively. According to this data the differences between the highest three treatments had no significance.

#### **Biomass dry weight(g):**

The data in Table (1) showed that the recommended dose (10:7.5:2.5g/pot) of 100% mineral NPK fertilizer T2 and Frankia (50g/pot) combined with 75% of mineral NPK fertilizer (7.5:3.75:1.875g) T8 followed by Mycorrhiza (50ml/pot) combined with 75% of mineral NPK fertilizer T10 produced The heaviest biomass dry weight of seedlings of casuarina with non significant differences between them whereas Minia Azotein (50ml/pot) combined with 75% of mineral NPK fertilizer T6 ranked the fourth in this concern

while the other treatments did not attain any significant variens in biomass dry weight compared to control (00.0)T1.

The increase in vegetative characteristics i.e. plant height, diameter, number of branches, biomass fresh weight, biomass dry weight fertilizer NPK are in parallel with those obtained by Abo El-Wafa (2014) and Abdou et al., (2014) on *Populus nigra* mentioned that fertilizing seedling with NPK at a rate 4:2:2 g/ pot improved its vegetative characteristics. On *Swietenia mahagoni* seedlings, Youssef et al. (2019) evaluated the effect of different NPK fertilizer rates (0, 4, and 6 g/plant) on vegetative growth. Results showed that clay media plus 6 gm NPK gave the highest values of plant height, leaves fresh and dry weights, and leaves number /plant. While, clay and sand (1:1) media plus 6 gm NPK recorded the highest values of leaves number/ plant compared with other treatments. The stimulating effect of chemical fertilization on Vegetative growth parameters It may be due to the role of chemical fertilization in supplying plants with the nutrients they need for production more carbohydrate and protein necessary for the vegetative growth of casuarina seedlings Vasanthakrishna, et al. (1994) on *Casuarina equisetifolia* Jihane et al. (2016) on *Casuarina* spp Frosi et al. (2016). On *Poincianella pyramidalis*, and *Cnidioscolus quercifolius* Abdou and Badran (2018) on *Delonix regia*, Kohan et al. (2000) and Abdou et al. (2014) on *Populus nigra*, Abd El-Aziz (2000) on *Azadirachta indica*, El-Mahrouk (2000) and Youssef et al. (2019) on *Swietenia* spp. Sayed (2001), Abdou et al. (2006) and Agera et al. (2019) on *Khaya senegalensis*, Abd El-Dayem (2003) on *Taxodium distichum*, Al-Menaie et al. (2010) on *Cassia nodosa* and *Cassia fistula*, Adejobi et al. (2015) on cocoa, Saravanakumar and Shanthinipriya (2017) on *Populus deltoids*. The increase in vegetative characteristics due to Chemical fertilizer NPK is in parallel with those obtained by Vasanthakrishna et al. (1994) on *Casuarina equisetifolia* and Sanginga et al. (1989) on *Allocasuarina* (*A. torulosa* and *A. littoralis*) and *Casuarina equisetifolia* since they found that the effect of phosphorous on seedling growth varies with plant species and whether nitrogen was derived by symbiotic fixation or given nitrate. The effects of nitrogen treatments (pollinating plants and plants) were fertilized with nitrogen) and phosphorous on the growth of *C. equisetifolia* significantly between 0 and 30 mg/kg of soil. Increasing the phosphorous level in the soil from 0 to 30 mg/kg led to an increase in the vegetative characteristics.

The increase in vegetative characteristics due to Minia Azotein fertilizer was deduced by Badran et al., (2003) on *Acacia* spp and Abo El-Wafa (2014) on *Populus nigra* mentioned that fertilizing Plants with Minia Azotein at a rate 50ml/pot improved its vegetative characteristics.

The mentioned results of vegetative characteristics concerning *Frankia* are coincided with those obtained by Saravanan et al. (2012) on *Casurina equistifolia* since fertilizing *Casurina equistifolia* plants with *Frankia* at a rate of 50 g/ seedling improved its vegetative characteristics. The results of bio-fertilizer with *Frankia* are in parallel with those obtained by Sanginga et al. (1989). In addition, seedlings of *C. equisetifolia* inoculated with *Frankia* had 7-10 times increase in the seedlings shoot (Karthieyan et al., 2012). They found that the inoculated *Casuarina equisetifolia* seedlings increased in vegetative growth parameters by between 90% -100% to the seedlings not inoculated with *Frankia*. The stimulating effect of bio-fertilizer with *Frankia* on vegetative growth parameters is due to its ability to convert atmospheric nitrogen into ammonia via the

enzyme nitrogenase, a process known as nitrogen fixation. They do this while living in root nodules on actinic plants. Bacteria can provide most or all of the nitrogen requirements of the host plant. As a result, actinic plants colonize and often thrive in soils low in phytonutrients (Sanginga et al., 1989). Similar trends were obtained by several authors like Diem et al. (1982), Vasanthakrishna et al. (1994) on *Casuarina equisetifolia*, Wheeler et al. (2000) on four species of casuarinas *C. cunninghamiana*, *C. equisetifolia*, *C. glauca*, *C. junghuniana*.

The increase in vegetative characteristics due to Mycorrhiza fertilizer was deduced by Abo EL-Leleil (2016) on *Swieteiana mahagony* mentioned that fertilizing Plants with Mycorrhiza at a rate of 150 spores improved its vegetative characteristics. The stimulating effect of bio-fertilizer with mycorrhizal on vegetative growth parameters may be due to the successful fungal symbiosis with the plant where the plant is a source of energy for the fungus in the form of fixed carbon, while the fungus gives nutrients to plants, the most important of which is phosphorous, which is present in the soil solution in very low concentrations as mentioned by Hayman (1983), Rajendran and Devaraj (2004) on *Casuarina equisetifolia*, Jihane et al. (2016) on *Casuarina* spp, Trappe (1982) on many important forest tree species, such as *liquidambar* (sweetgum), *fraxinus* (ash), *liriodendron* (poplar), Schmitz-zeitz (1995) on *Fagus sylvatica*. The results of biofertilizer with mycorrhizal fungi are parallel to those obtained by Karthieyan et al. (2012) and they found that the inoculated seedlings increased in vegetative characteristics by between 90%-100% over the non-inoculated seedlings.

#### Root growth characteristics

##### Root length (cm):

The data in Table (1) showed a significant increase in root length of *Casuarina equisetifolia* seedlings compared to control. The longest root length were recorded when mineral NPK fertilizer as a recommended dose (10:7.5:2.5g/pot) was used and attained the highly percentages of increases reached to 105.26 and 77.77 % over control in the first and second season respectively. Followed by treatment 75% of NPK with *Frankia* (50g/plant) which gave a significant increase in root length reached to 101.31 and 74.49 % over control in the first and second season respectively, then treatment of Mycorrhiza (50ml/pot) plus 75% NPK, which gave a significant increase in length reached to 83.94 and 72.22 % over control in the first and second season respectively. According to this data the differences between the highest three treatments had no significance.

##### Root fresh weight (g):

The data in Table (2) showed that the recommended dose (10:7.5:2.5g/pot) of 100% mineral NPK fertilizer T2 and *Frankia* (50g/pot) combined with 75% of mineral NPK fertilizer (7.5:3.75:1.875g) T8 followed by Mycorrhiza (50ml/pot) combined with 75% of mineral NPK fertilizer T10 produced The heaviest root fresh weight of seedlings of casuarina with non significant differences between them whereas Minia Azotein (50ml/pot) combined with 75% of mineral NPK fertilizer T6 ranked the fourth in this concern while the other treatments did not attain any significant variens in root fresh weight compared to control (00.0)T1.

##### Root dry weight (g):

The data in Table (1) showed a significant increase in root dry weight compared to the control. The heaviest weight of

root dry weight was recorded when the mineral NPK fertilizer as a recommended dose was used and attained the high percentages of increases reached to 30.85 and 33.05 % over control in the first and second season respectively. Followed by treatment 75% of NPK with Frankia (50g/plant) which gave a significant increase in root dry weight reached to 29.88 and 31.33 % over control in the first and second season respectively, then treatment by Mycorrhiza (50ml/pot) plus 75% NPK, which gave a significant increase in root dry weight reached to 29.67 and 29.92 % over control in the first and second season respectively. According to this data, the differences between the highest three treatments had no significance.

The increase in root growth characteristics i.e. root length, root fresh and dry weight due to NPK fertilizer are in parallel with those obtained by Abo El-Wafa (2014) and Abdou *et al.* (2014) on *Populus nigra* who mentioned that fertilizing seedlings with NPK at a rate 4:2:2 g/ pot improved its root characteristics.

The stimulating effect of chemical fertilization on root growth characteristics may be due to the role of chemical fertilization in supplying plants with the nutrients they need for production more carbohydrate and protein necessary for the root growth of casuarina seedlings as mentioned by several authors like Vasanthakrishna *et al.* (1994) on *Casuarina equisetifolia* Jihane *et al.* (2016) on *Casuarina spp* Frosi *et al.* (2016) on *Poincianella pyramidalis*, and *Cnidocolus quercifolius* Abdou and Badran (2018) on *Delonix regia*, Kohan *et al.* (2000) and Abdou *et al.* (2014) on *Populus nigra*, Abd El-Aziz (2000) on *Azadirachta indica*, El-Mahrouk (2000) and Youssef *et al.* (2019) on *Swietenia spp.* Sayed (2001), Abdou *et al.* (2006) and Agera *et al.* (2019) on *Khaya senegalensis*, Abd El-Dayem (2003) on *Taxodium distichum*, Al-Menaie *et al.* (2010) on *Cassia nodosa* and *Cassia fistula*, Adejobi *et al.* (2015) on cocoa, Saravanakumar and Shanthinipriya (2017) on *Populus deltoids*.

The results of chemical fertilization are in parallel with those obtained by Vasanthakrishna *et al.* (1994) on *Casuarina equisetifolia*. Likewise, Sanginga *et al.* (1989) work on *Allocasuarina (A. torulosa* and *A. littoralis)* and *Casuarina equisetifolia* and found that the effect of phosphorous on seedling growth varies with plant species and whether nitrogen was derived by symbiotic fixation or given nitrate. The effects of nitrogen treatments (pollinating plants and plants) were fertilized with nitrogen) and phosphorous on the growth of *C.*

*equisetifolia* significantly between 0 and 30 mg/kg of soil. Increasing the phosphorous level in the soil from 0 to 30 mg/kg led to an increase in the dry weight of the plant and the dry weight of the roots.

The increase in root characteristics due to Mycorrhiza fertilizer was deduced by Abo EL- Leleil (2016) on *Swietenia mahagony* mentioned that fertilizing plants with Mycorrhiza at a rate (150spores) improved its root characteristics. The stimulating effect of bio-fertilizer with Mycorrhiza on root characteristics parameters It is due to the successful fungal symbiosis with the plant where the plant is a source of energy for the fungus in the form of fixed carbon, while the fungus gives nutrients to plants, the most important of which is phosphorous, which is present in the soil solution in very low concentrations (Hayman, 1983). Rajendran and Devaraj (2004) on *Casuarina equisetifolia*, Jihane *et al.* (2016) on *Casuarina spp*, Trappe (1982). on many important forest tree species, such as *liquidambar* (sweetgum), *fraxinus* (ash), and *liriodendron* (poplar), Schmitz-zeitz (1995). on *Fagus sylvatica* were in the same way.

The mentioned results of root characteristics concerning Frankia coincide with those obtained by Saravanan *et al.* (2012) on *Casuarina aquistifolia* who revealed that fertilizing *Casuarina equistifolia* plants with Frankia at a rate of 50 g/seedling improved its root characteristics. The results of bio-fertilizer with Frankia are in parallel with those obtained by Sanginga *et al.* (1989) who found that seedlings of *C. equisetifolia* inoculated with Frankia had 7-10 times the dry weight of un inoculated seedlings in terms of shoot and root dry weight. Karthiyan *et al.* (2012) found that the inoculated *Casuarina equisetifolia* seedlings increased in root characteristics parameters by between 90% -100% than the seedlings not inoculated with Frankia. The stimulating effect of bio-fertilizer with Frankia on root characteristics parameters may be due to its ability to convert atmospheric nitrogen into ammonia via the enzyme nitrogenase, a process known as nitrogen fixation. They do this while living in root nodules on actinic plants. Bacteria can provide most or all of the nitrogen requirements of the host plant. As a result, actinic plants colonize and often thrive in soils low in phytonutrients (Sanginga *et al.*, 1989). Our findings were supported by those which obtained by Diem *et al.* (1982), Vasanthakrishna *et al.* (1994) on *Casuarina equisetifolia*, Wheeler *et al.* (2000) on four species of casuarinas *C. cunninghamiana*, *C. equisetifolia*, *C. glauca*, *C. junghuniana*

**Table 1. Effect of biofertilizers (Minia Azotein (MA) , Frankia and Vesicular Arbuscular Mycorrhiza (VAM) and chemical fertilizer NPK on Plant height (cm), Stem diameter (cm), No. of branches, Biomass fresh & dry weights (g) of *Casuarina aquisetifolia* seedlings in both seasons 2019 and 2020.**

Treatments	Plant height (cm)		Stem diameter (cm)		No. of branches		Biomass fresh weight (g)		Biomass dry weight (g)	
	1 <sup>st</sup> season	2 <sup>nd</sup> season	1 <sup>st</sup> season	2 <sup>nd</sup> season	1 <sup>st</sup> season	2 <sup>nd</sup> season	1 <sup>st</sup> season	2 <sup>nd</sup> season	1 <sup>st</sup> season	2 <sup>nd</sup> season
T1	122.0	129.1	1.17	1.53	36.1	36.4	469.7	481.25	140.80	148.94
T2	180.3	195.3	2.27	2.33	49.7	51.4	695.8	717.04	258.80	273.75
T3	128.5	130.0	1.26	1.61	36.2	38.2	471.7	483.34	141.80	149.99
T4	133.7	130.5	1.37	1.77	36.3	40.3	472.8	484.48	142.30	150.52
T5	133.7	130.5	1.37	1.77	36.3	40.3	472.8	484.48	142.30	150.52
T6	165.5	180.3	1.93	2.18	44.4	47.2	607.5	624.95	212.70	224.99
T7	144.9	160.5	1.64	1.93	40.8	43.5	535.4	553.12	175.10	185.22
T8	179.3	195.0	2.20	2.30	48.9	50.3	682.8	703.48	252.00	266.56
T9	159.3	166.8	1.81	2.06	42.7	44.9	548.6	563.53	181.90	192.41
T10	174.6	189.0	2.18	2.26	48.3	49.6	673.0	693.26	246.80	261.06
T11	150.6	166.1	1.75	2.01	41.1	44.2	547.1	561.97	181.20	191.67
L.S.D. at 5%	5.7	6.5	0.12	0.11	1.4	1.8	51.5	53.5	16.8	18.2

T1=Control, T2=100 % NPK, T3= Minia Azotein at 50 ml/plant, T4= Frankia at 50 g /plant, T5= Mycorrhiza at 50 ml /plant, T6= 75 % NPK+ Minia Azotein, T7=50 % NPK+ Minia Azotein, T8=75 % NPK+ Frankia, T9=50 % NPK+ Frankia, T10=75 % NPK+ Mycorrhiza, T11=50 % NPK+ Mycorrhiza

**Table 2. Effect of biofertilizers Minia Azotein (MA), Frankia, Vesicular Arbuscular Mycorrhiza (VAM), and chemical fertilizer NPK on root length(cm), root fresh and dry weights(g) of *Casuarina equisetifolia* seedlings in both seasons 2019 and 2020.**

Treatments	Root length(cm)		Root fresh weight (g)		Root dry weight(g)	
	1 <sup>st</sup> season	2 <sup>nd</sup> season	1 <sup>st</sup> season	2 <sup>nd</sup> season	1 <sup>st</sup> season	2 <sup>nd</sup> season
T1	38.00	39.6	247.22	248.00	137.14	137.42
T2	78.00	70.4	363.85	373.18	179.46	182.85
T3	38.30	45.2	254.00	255.15	139.60	140.01
T4	47.30	48.6	271.44	282.51	145.93	149.94
T5	38.50	46.2	260.17	270.14	141.84	145.45
T6	66.70	57.9	321.28	323.57	164.01	164.84
T7	48.90	50.0	283.37	284.76	150.25	150.76
T8	76.50	69.1	360.16	366.42	178.12	180.39
T9	57.50	53.1	309.38	311.31	159.69	160.39
T10	69.9	68.2	359.36	361.31	177.83	178.54
T11	49.20	52.8	297.57	299.06	155.41	155.95
L.S.D. at 5%	8.4	2.3	11.22	12.11	4.05	4.35

T1=Control T2=100 % NPK T3= Minia Azotein at 50 ml/plant T4= Frankia at 50 g /plant T5= Mycorrhiza at 50 ml /plant T6= 75 % NPK+ Minia Azotein T7=50 % NPK+ Minia Azotein T8=75 % NPK+ Frankia T9=50 % NPK+ Frankia T10=75 % NPK+ Mycorrhiza T11=50 % NPK+ Mycorrhiza

Mycorrhizal Fungi play a role in the process of plant growth as a result of their secretion of some organic compounds and the intertwining of their cells with the roots of the host, which helps to stabilize the soil granules, and increase its ability to retain water, which helps to increase agricultural production (Barrios, 2007; Smith and Read, 2008). VAM inoculation causes an increase in the vegetative characteristics and production of various crops, and it works to improve the properties of sandy soil and increase its content of nitrogen, phosphorous, and potassium. It also works to increase the amount of humus and thus expands its ability to retain water.

Frankia strains are nitrogen-fixing actinomycetes that were isolated for the first time in 1978. They form root nodules. The vesicles contain the enzyme nitrogenase, which is important in the process of atmospheric nitrogen fixation, which helps to solve many soil problems and increase reclaimed areas. (Fontaine *et al.*, 1986; Sayed *et al.*, 2000; Hahn *et al.*, 2003). Frankia inoculation causes an increase in the vegetative characteristics and production of various crops and it works to improve the properties of sandy soil and increase its content of nitrogen, phosphorous, and potassium. It also works to increase the amount of humus and thus expands its ability to retain water.

Chemical fertilizers enter most of the processes that enter the plant, where nitrogen enters the components of most organic compounds such as nucleic acids (RNA and DNA), amino acids, alkaloids, vitamins, and enzymes. Phosphorous is necessary for plants as it enters as one of the components of the cell nucleus and is of great importance in cell division and development of meristem cells and enters the formation of phospholipids nucleic acids and has an important role in the phosphorylation process that causes the production of energy compounds (ADP and ATP). Most of the physiological processes, such as carbohydrate formation, photosynthesis and other various processes within the plant. Potassium is necessary for most of the physiological processes that take place inside the plant, as it has an important role in the process of nitrogen metabolism, and has a role in the work of enzymes. On the osmotic pressure of cells Ingels (1994), Yagodin (1984), Mengel and Kirkby (1987) and Bhuiyan, *et al.* (2000). Mineral fertilization causes an increase in the chemical characteristics and production of various crops. The stimulating effect of chemical fertilization may be due to the role of chemicals in supplying the plants with their required

nutrients for more carbohydrates and protein production which are necessary for the increase in total chlorophyll contents of *Casuarina equisetifolia* seedlings. Sanginga *et al.* (1989) on *Allocasuarina* (*A. torulosa* and *A. littoralis*) and *Casuarina equisetifolia*. Danso (1990) on *Casuarina equisetifolia*. and Vasanthakrishna *et al.* (1994) on *Casuarina equisetifolia*.

Finally, the results showed that all treated seedlings of casuarina with frankia, Vesicular Arbuscular Mycorrhiza (VAM) in combination with 75% NPK (T8 and T10) and or NPK, 100% recommended dose (T2) recorded significant increases in vegetative characteristics i.e. Plant height, Stem diameter, Number of branches, Biomass fresh and dry weights compared to the other treatments and control. The highest rooting values of root length, root fresh, and dry weights of casuarina plants were found with the abovementioned treatments followed by Minia Azotein (MA) combined with 75%NPK (T6) in both seasons. while the other treatments did not attain any significant variances in vegetative and root growth parameters. It can be concluded that Frankia 50g/plant, Vesicular Arbuscular Mycorrhiza (VAM) 50 ml/plant with 75% of the recommended dose (7.5:3:75.5:1.875g/plant)T6, T8, and/ or 100% of the recommended dose of NPK (10:5:2.5g/plant) T2 were the best in improving vegetative and root characteristics of *Casuarina equisetifolia* seedlings.

## CONCLUSION

From the results, it is recommended to fertilize *Casuarina equisetifolia* seedlings grown in a pot of sandy soil with 7.5 g of nitrogen fertilizer (ammonium sulfate 20.5% N), 3.75 g of phosphate fertilizer (calcium superphosphate 15.5%) P<sub>2</sub>O<sub>5</sub> and 1.875 g of potassium fertilizer (potassium sulfate 48 % K<sub>2</sub>O) with the addition of one of the biofertilizers (Frankia at 50 g / seedling or Mycorrhiza at 50 ml/seedling in order to decline 25% of chemical fertilizer and to improve the vegetative and root growth of *Casuarina equisetifolia* seedlings.

## REFERENCES

- Abd El-Aziz, M.F. (2000): Effect of soil types and NPK fertilization treatments on *Azadirachta indica* seedlings, M.Sc. Thesis, Fac. of Agric., Minia Univ.
- Abd El-Dayem, A.M. (2003): Effect of fertilizer treatments in *Taxodium distichum* seedlings grown in alkali soil. For. Bce Manage. 33134: 531-541 (C.F. Tree Physiol. 16; 307-313).

- Abdou, M. A. H.; Ahmed, E.T.; Ahmed, A. A. and Abdel-Mola, M. A. M. (2014): Response of *Populus nigra*, l. seedlings to compost, biofertilizers and mineral NPK fertilization. *Minia J. of Agric. Res. & Develop.* Vol. (34), No. 1, pp. 31-47, 2014.
- Abdou, M.A.H. and Badran, F.S. (2018): Effect of NPK fertilization and micronutrients on *Delonix regia* seedlings grown in sandy soil. *Scientific J. Flowers & Ornamental Plants*, 5(2):151-159.
- Abdou, M.A.H.; Ahmed, E.T.; Taha, R.A. and Helmy, S.M.S. (2006): Response of *Khaya senegalensis* to some bio and chemical fertilization treatments. *Minia J. of Agric. Res. & Develop.*
- Abo El-Wafa, M.K. (2014): Physiological studies on *Populus nigra* seedlings. M.Sc. Thesis, Fac. of Agric., Minia Univ.
- Abou El-Ghait, E.M.; Youssef, A.S.M.; Mohamed, Y.F.; Noor El-Deen, T.M. and Heba, I. Mohamed (2020). Effect of benzyladenine and chemical fertilization on growth, flowering and chemical composition of Jasmine sambac plant. *Scientific J. Flowers & Ornamental Plants*, 7(4): 1-13.
- Abo EL- Leleil, N.K. A. (2016): Effect of sewage, sludge, mineral fertilization and endomycorrhizal fungi on growth of *Swietenia mahagoni* seedlings under soil conditions. M.Sc. Thesis, Fac. of Agric., Zagazig Univ.
- Adejobi, K.B.; Akanbi, O.S.; Ugiori and (ed.) (2015): Comparative effects of NPK fertilizer, cowpea pod husk and some tree crops wastes on soil, leaf chemical properties and growth performance of cocoa (*Theobroma cacao*, L.). *African Journal of plant science*, 8(2): 103-107.
- Agera, S.I.N.; Amonum, J.I. and Kuje, E.D. (2019): Effect of varying levels of fertilizer and organic manure on growth of *Khaya senegalensis* seedlings in Benue State, North Central Nigeria. *Res. J. Agriculture and Forestry Sci.*, Vol. 7(2), 1-9, April (2019).
- Al-Menaie, H.S.; Al-Ragon, O. ; Al-Shatti, M.; Mathew, M. and Suresh, N. (2010) : Evaluating the growth performance of *Cassia nodosa* and *Cassia fistula*, L. seedlings using different potting mixtures. *Academic J. of Plant Sci.*, 3 (1) : 33-36.
- Alok, C.(2021). Toru Dutt's "Our Casuarina Tree": An Exploration of Its Significance to Global Ecological Harmony. *Journal literary Herald* Vol. ( 6) 173-179.
- Anthony, D.; Glass, M. (2010). Nitrogen Use Efficiency of Crop Plants: Physiological Constraints upon Nitrogen Absorption. *Journal plant sciences*. Vol 22(5).453-470.
- Arnold, J. B. (2015). The increasing importance of distinguishing among plant nitrogen sources. *Journal Science Direct*. (25)10–16 [www.sciencedirect.com](http://www.sciencedirect.com).
- Badr El-Din, S.M.S.; Attia, M. and AboSedera, S.A. (1999). Evaluation of several substrates for mass multiplication of arbuscular mycorrhizal (AM) fungi grown on onion. *Egyptian Journal of Microbiology*, 34:57-61.
- Badran, F.S.; Abdou, M.A.; Aly, M.K.; Sharaf, El-Deen, M.N. and Mohamed, S.H. (2003): Response of sandy soil grown *Acacia saligna* seedlings to organic bio-and chemical fertilization and IAA treatments. 1st Egyptian Syrian Conf. for Agric. and Food in the Arabian Nation, Minia Univ., Minia, Egypt, December, 8–11.
- Barrios, E. (2007). Soil biota, ecosystem services and land productivity. *Ecological Economics*, 64:269-285.
- Bhuiyan, M. Z. A.; Hossain, M. K.; Osman, K. T.) (2000) ( Effect of inorganic fertilizers on the initial growth performance of *Casuarina equisetifolia* seedlings in the nursery. *Indian Journal of Forestry* Vol.23 No.3 pp.296-300 ref.29
- Crocker, L.J. and Schwintzer, C.R. (1993): Factors affecting formation of cluster roots in *Myrica gale* seedlings in water culture. *Plant and Soil*, 152(2): 287-298.
- Curtis, P.S. (1996). A meta-analysis of leaf gas exchange and nitrogen in trees grown under elevated carbon dioxide. *Journal Plant, cell and Environment*. Vol 19(2).127-137.
- Dadarwall, L.R.; Yadv, L.S. and Sindhu, S.S. (1997): Biofertilizer production: Technology: Prospects in biotechnological approach in soil microorganisms for sustainable crop production. *Scientific Publishers, Jodhpur., India*. P. 323 – 337.
- Danso (1990). Estimation of N<sub>2</sub> and N fixation is derived from Soil by *Casuarina EQUSETIFOLIA* using stickers "N Fertilizer: Some Problems and Solutions. *Soil Bd. B&hem*. Vol. 22. No. 5. pp. 695-701.
- Diagne, N.; Diouf, D.; Svistoonoff, S.; Kane, A.; Noba, K.; Franche, C.; Bogusz, D. and Duponnois, R. (2013): *Casuarina* in Africa: distribution, role and importance of arbuscular mycorrhizal, ectomycorrhizal fungi and Frankia on plant development. *J. Environ Manage*, 128:204-209.
- Diem, H. G.; Gauthier, D. and Dommergues Y.R. (1982). Extranodular growth of Frankia on *Casuarina equisetifolia*., *FEMS Microbiology Letters* (1982) 15 181-184
- Dinkelaker, B.; Hengeler, C. and Marschner, H. (1995): Distribution and function of proteoid roots and other root clusters. *Bot Acta*, 108(3): 183–200.
- El-Mahrouk, E.M. (2000): Response of *Swietenia macrophylla*, L. to different levels of irrigation water and NPK fertilization treatments in a newly reclaimed area. *J. Agric. Res., Tanta Univ.*, 26 (2): 377 – 390.
- Frosi, G.; Barrosa, V.A.; Oliveiraa, M.T.; Cavalcanteb U.M.T.; Maiab, L.C.; and santosa, M.G. (2016): Increase in biomass of tow woody species from a seasonal dry tropical forest in association with AMF with different phosphorus levels. *Applied Soil Ecology* 102(2016)46-52.
- Fontaine, M.S.; Young, P.H.; Torry, J.G. (1986). Effects of long-term preservation of Frankia strains on infectivity, effectivity, and in vitro nitrogenase activity. *Applied Environmental Microbiology* 51: 694-698.
- Ganaw, H.A.E. (2017): Effect of bio and chemical fertilization and planting media on growth and chemical composition of *Moringa oleifera* trees. Ph.D. Thesis, Fac. of Agric., Cairo Univ.
- Hayman, D. S. (1983). The physiology of vesicular- arbuscular endomycorrhizal symbiosis. *Can. J. Bot.*, 61:944-963.
- Hauwaka, F.I.A. (2000) : Effect of using single and composite inoculation with *Azospirillum brasilense*, *Bacillus megatherium* var. *phosphaticum* and *Glomus marcocarpus* for improving growth of *Zea mays*. *J. Agric. Sci. Mansoura Univ.*, 25 (1) : 239 – 252..

- Hahn, A.B.; Hock, M.M. Animon, R.; Naryanan,; and Wheeler C.T. (2003).The production and utilization of monoclonal antibodies for identification of a Frankia utilized as inoculum for Casuarina equisetifolia. Plant and Soil 254: 27-33.
- Helly , C.; Ponnusamy ,S. K.; Muthulingam, S.; MadhavaAnil, K.(2021). Kinetics, equilibrium and thermodynamic investigations of methylene blue dye removal using *Casuarina equisetifolia* pines. Journal science direct vol( 285).131-150.
- Hedge, D.M.; Dwivedi, B.S. and Sudhakara, B.S.S. (1999) : Biofertilizers for cereal production in India – A review. Indian J. Agric. Res., 69 (2) : 73 – 83.
- Ian T.Rley (2021). A case for assessing allocasuarina and casuarina SSP. For use in agroecosystem improvement in semi-areas with a focus on central Anatolia turkey. Journal Front. Agr. Sci. Eng. 8(4): 568–582.
- Ingels, J.E. (1994) : Ornamental Horticultural Science, Operation & Management. 2nd Edition. ITP Delmar Publishers Inc.
- James, A.R. and Michael, R.E. (2009): Growing NPK media for container production in green house or nursery.Agriculture and Natural Resources. <http://www.uaex.edu>.
- James, J.E. (2020) . Phosphorus: a limiting nutrient for humanity. Journal Biotechnology.(23).6.833-838.
- Jihane Touati, Mohamed Chliyeh, Amina Ouazzani Touhami, Rachid Benkirane and Allal Douira (2016): Effect of mycorrhizae on growth and root development of Casuarina spp. under greenhouse conditions. J. IJAPBC – Vol. 5(3), 2016 ISSN: 2277 – 4688. [www.ijapbc.com](http://www.ijapbc.com)
- Kaan, I.; Ramazan, E.(2022). The effect of atmospheric deposition on potassium accumulation in several tree species as a biomonitor. Journal Environmental Research and Technology.vol (5).(1).94-100.
- Karthikeyan, A.; and Krishnakumar, N. (2012). Reforestation of bauxite mine spoils with Eucalyptus tereticornis Sm. seedlings inoculated with arbuscular mycorrhizal fungi. Ann. For. Res. 55(2): 207-216.
- Karthikeyan, A. (2016): Frankia strains for improving growth, biomass and nitrogen fixation in *Casuarina equisetifolia* seedlings. Journal of Tropical Forest Science 28(3): 235-242 .
- Kohan, S.; Kosice, S. and Vyskumna, S. (2000): Effect of fertilizing on the production and health of poplar 1-214 (*Populus × Euramericana* cv. 1-214) in intensive plantations. J. For. Sci., 46 (7): 325 – 330.
- Mackay, J.; Simon, L. and Lalonde, M.(1987). Effect of substrate nitrogen on the performance of in vitro propagated *Alnus glutinosa* clones inoculated with Sp+ and Sp– Frankia strains. Journal Plant and Soil.vol (103).21-31.
- Malajczuck, N. and Lamont, B.B. (1981): Specialized roots of symbiotic origin in heath lands. In: Heath lands and Related Shrublands of the Worl, B. Analytical Studies. Specht R.L. (Ed.), Elsevier, Amsterdam, 1981; 165-182.
- Masindi, V., Muedi, K. (2018). Environmental contamination by heavy metals, heavy metals. Book Heavy Metals.p77-90.
- Mengel, K . and Kirkby, A. (1987): Principles of plant nutrition. 4th Ed International Potash, Institute, Bern/ Switzerland
- Murry M.A.: M.S Fontaine and J.G. Torrey (1984): Growth kinetics and nitrogenase induction in Frankia sp.HFPAr13 grown in batch culture . plant and soil 78:61-78
- MSTAT–C (1986): A microcomputer program for the design management and analysis of Agronomic Research Experiments (version 4.0), Michigan State Univ., U.S.A.
- Nikita, B. and Puneet, S. (2020). Excessive and Disproportionate Use of Chemicals Cause Soil Contamination and Nutritional Stress. Book Soil Contamination.p5-18.
- Ojha, S.; and Arya, A. (2018): Increase in plant growth and biomass of *Casuarina equisetifolia*, L. by incorporating three different fungi in the rhizosphere. Current Research in Environmental & Applied Mycology (Journal of Fungal Biology) 8(1):96–103(2018) [www.creamjournal.org](http://www.creamjournal.org)
- Okalebo JR, Kenneth WG and Woome, P.L. (2002): Laboratory method of soil and plant analysis. Second edition, SACRED Office Nairobi, p 128.
- Pooja, J. M. and Katharina, P. (2021): Anthropogenic influences on the distribution of the *Casuarina-Frankia* symbiosis. Symbiosis 84:353–367.
- Rashed, A. (2017): Effect of some agricultural treatments on growth, yield and chemical constituents of moringa seedlings. M.Sc. Thesis, Fac. of Agric. Al-Azhar Univ., Assiut.
- Rajendran, K. and Devaraj, P. (2004): Biomass and nutrient distribution and their return of *Casuarina equisetifolia* inoculated with biofertilizers in farm land. Biomass and Bioenergy. (3)235-249
- Sanginga, N.; Danso S.K.A.; Bowen G.D. (1989). Nodulation and growth response of *Allocasuarina* and *Casuarina equisetifolia* to phosphorus fertilization. Plant and Soil 118, 125-132.
- Saravanakumar, R. and Shanthinipriya, A. (2017): Effect of inorganic fertilizers and growth performance in poplar tree *Populus deltoids*, Tamilnadu, India. Res. J. Agriculture and Forestry Sci., Vol. 5(9), 13-17.
- Saravanan, T.S.; Rajendran, K. and Santhaguru K.(2012). Selection of Suitable Biofertilizers for Production of Quality Seedlings of Casuarina Equisetifolia (Forst.) Using Decomposed Coir Pith Compost in Root Trainers. Journal ASIAN J.EXP.BIOL.SCI 3(4).752-761.
- Sayed, R.M. (2001): Effect of some agricultural treatments on the growth and chemical composition of some woody tree seedlings. Ph.D. Thesis, Fac. Agric., Minia Univ.
- Sayed, W.F.; Mohawad,S.M .and Abd El-karim,M.M.(2000). Effect of Al,Co,and Pb ions on growth of Frankia sp. in a mineral medium. Folia Microbiologica 45: 153-156.
- Schmitz-Zeitz, D. (1995): Use of mycorrhizal in a long –term experiment. AFZ / Der Wald, Allgemeine forest Zeitschrift furb Waldwirtschaft and Umweltvorsorge; 50(23):1264-1256. 7 ref.

- Shantharam, S. and Mattoo, A.K. (1997): Enhancing biological nitrogen fixation: an appraisal of amount and alternative technologies for N in put into plants. *Plant Soil* 194 205–216.
- Smith, S.E. and Read, D. (2008). *Mycorrhizal symbiosis*. Academic Press, London, UK.
- Soliman, Sh.H.H. (2019): Effect of some agricultural treatments on *Moringa peregrina* plant. Ph. D. Thesis, Fac. of Agric., Minia Univ.
- Trappe, J.M. (1982). Synoptic keys to the genera and species of zygomycetous mycorrhizal fungi. *Phytopathology*, 72(8): 1102\_1108
- Vain, M.; Latha S. K.; Ratna Harika C.; Tejaswi K. S.; Sareddu, S. Pattipati, P.(2022). A detailed investigation of phytochemical, biological and commercial utilization of horse tail tree *Casuarina equisetifolia*. *Asian Journal of Pharmaceutical Research*. Vol 12, Issue - 1, 1-7
- Vasanthakrishna, M.; Bagyaraj, D.J.; and Nirmalnath, P.J.(1994): Responses of *Casuarina equisetifolia* to inoculation with *Glomus fasciculatum* and/or *Frankia*. *Forest Ecology and Management* 68 (1994) pp399-402
- Wei, Xiao.; Xifei, Li.; Bin, Cao.; Gang, Huang.; Chong, Xie.; Jian, Qin.; Huijuan, Yangab.; Jingjing, Wangab. and Xueliang, Sun.(2021). Constructing high-rate and long-life phosphorus/carbon anodes for potassium-ion batteries through rational nanoconfinement. *Journal Nano Energy*.vol 38.1-14.
- Wheeler, C.T.; Tilak, M.; Scrimgeour, C.M.; Hooker, J.E. and Handley L.L. (2000): The effects of symbiosis with *Frankia* on the natural abundance of N<sub>15</sub> in four species of casuarinas. *Journal of Experimental Botany*, Vol.51 No.34, PP.287-297.
- Wheeler, G. S.; Taylor, G. S.; Gaskin, J. F.(2011). Ecology and Management of Sheoak (*Casuarina* spp.), an Invader of Coastal Florida, U.S.A. *Journal of Coastal Research* 27 (3): 485–492.
- Yagodin, B. A. (ed). (1984): *Agricultural Chemistry Part II*. Mir Publishers, Moscow. pp 1-66.
- Yang, Y.; Xinyu, Z.; Iain, P. H.; Jennifer A. J. D.; Xuefa, W.; Dandan, L.; Zhiming, G. and Timothy, A. Q.(2022). Contrasting rhizosphere soil nutrient economy of plants associated with arbuscular mycorrhizal and ectomycorrhizal fungi in karst forests. *Journal Plant Soil*. Vol 470.p81-93.
- Yongcheng, W.; Zhang, J.; Meng, Y.; Wang, C.; Zhong, H.(2021). Transcriptome and metabolome profiling in naturally infested *Casuarina equisetifolia* clones by *Ralstonia solanacearum*. *Journal Genomics* 113, 1906–1918.
- Youssef, N.M.; EL Habba, E.; Abd El- Aziz, N.G.; Mazhar, A. A.; Sarhaan, A.M.Z. and Arafa, A.M.S. (2019): Influence of using NPK fertilizer on vegetative growth and nutrients content of *Swietenia mahagoni*, (L.) Jacq. plants growing in different media types. *Curr. Sci. Int.*, 8(1): 62-69.
- Yujin, ;Yingting, X.; Zhengwan, H.; Zhongyu, Z.; Xiaoyi, W.(2021). Metabolite pattern in root nodules of the actinorhizal plant *Casuarina equisetifolia*. *Journal Phytochemistry*. Vol 186.17-28.

## تأثير الاسمدة الحيوية والمعدنية على النمو الخضري والجذري لشتلات *Casuarina equisetifolia* النامية في الاراضي الرملية.

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القسم البساتين بالكلية. الزراعة ، جامعة بنها ، مصر  
القسم الاراضي بالكلية. الزراعة ، جامعة بنها ، مصر

أجريت هذه الدراسة في مزرعة ومختبرات قسم البساتين بكلية الزراعة بمشتهر ، جامعة بنها ، خلال موسمين متتاليين لعامي 2019 و 2020 على شتلات الكازورينا تمت الدراسة من خلال استخدام بعض الاسمدة الحيوية والاسمدة الكيماوية NPK بهدف تقييم فاعلية المنيا أزوتين MA والفرانكيا وفطريات الميكورايزا الجذرية (VAM) كلا بمفردها او بالتداخل بينهم على الصفات الخضرية والجذرية لشتلات الكازورينا المزروعة في تربة رملية . أظهرت النتائج زيادة في الصفات الخضرية محل الدراسة وهي ارتفاع الشتلة وسمكها وعدد فروعها وكتلتها الحيوية الطازجة والجافة وكذلك الصفات الجذرية وهي طول الجذر ووزنه الطازج والجاف مقارنة بالشتلات الغير معاملة خلال موسمي التجربة حيث أعطت الشتلات المسمدة بالجرعة الموصى بها من السماد الكيماوي NPK ( 2.5 : 5 : 10 / جم / اصيص ) أعلى قيمة في الصفات المذكورة أعلاها تلتها الشتلات المخصبة بالفرانكيا بمعدل ( 50 جم / للأصيص ) بالتداخل مع 75% من الجرعة الموصى بها من السمادي الكيماوي ( 1.875 : 3.75 : 7.5 ) كما سجلت الشتلات المخصبة بالميكورايزا بمعدل ( 50 مل / اصيص + 75% من الجرعة الموصى بها من NPK القيمة التالية بدون اي فروق معنوية بين المعاملات المذكورة اعلاها وتم الحصول على أقل القيم بواسطة الشتلة الغير معاملة باي من الاسمدة الكيماوية او الحيوية . وعليه يمكن التوصية بتسميد شتلات الكازورينا المنزرعة بالتربة الرملية بالتخصيب بالفرانكيا بمعدل ( 50 جرام / اصيص + 75% من الجرعة الموصى بها من NPK ) يمكن التوصية بالتخصيب بالميكورايزا بمعدل ( 50 مل / اصيص ) + 75% من الجرعة الموصى من NPK لتحسين الصفات الخضرية والجذرية لشتلات الكازورينا (*Casuarina equistifolia*).